# OBBR Office of Biorepositories and Biospecimen Research



# BRN Tissue Acquisition Variables Project

Helen M. Moore, PhD January 24, 2012





# Multiple pre-analytical factors can affect the molecular integrity of the biospecimen

BBR Office of Biorepositories and Biospecimen Research





# How Can Changes in Molecular Integrity of Biospecimens Affect Molecular Readout?

OBBR Office of Biorepositories and Biospecimen Research

# Genomics

**Proteomics** 

**Metabolomics** 



Changes in specific transcript levels based on ischemic time, not disease Lack of reproducibility of protein biomarkers in discovery research
Inconsistent IHC results in Research and Clinical Labs Inconsistencies in small molecule readouts, yielding results that point to the wrong pathway



Office of Biorepositories and Biospecimen Research

# FFPE: Delay to Fixation and Molecular Assay Results OBB

#### HER2 IHC and FISH in Breast Cancer: Loss of Biomarker Signal with Time to Fixation



Khoury T, et al., Mod Pathol. 2009 Nov;22(11):1457-67

Phosphoprotein pMAPK IHC of Colon Cancer : Gain of Biomarker Signal with Time to Fixation



Hartmut Juhl, Indivumed GmbH, BRN

# BRN Project in Tissue Acquisition and Processing Variables

OBBR Office of Biorepositories and Biospecimen Research



# **BRN Scientific Steering Committee and Subcommittees**

OBBR Office of Biorepositories and Biospecimen Research

- Interdisciplinary Committee
  - Pathologists, Statisticians, Scientists, Informaticist, Patient Advocate
- Working Group Subcommittees
  - IHC/FSH
  - DNA/RNA
- Subcommittees developed plans for:
  - Markers to be tested
  - Processes and procedures for testing markers
  - Data output desired
  - Data Analysis

# **Experimental Design Overview**

- Goal: to understand how variability in tissue fixation and processing affects the molecular integrity of the resulting tissues
- Approach: treat tissue fixation and processing (FFPE) as a manufacturing process which we want to study, but over which we do not have complete control
- Identify key preanalytical factors that expert intuition says are of most importance to getting a "good" biospecimen in the clinic
- Study these key factors in a first set of experiments to test expert intuition
- Evaluate experimental results and then plan the next set of experiments

# **Experimental Design Overview**

### OBBR Office of Biorepositories and Biospecimen Research

• 2 key preanalytical factors identified for first set of experiments:

- Delay time to fixation
- Time in fixative before processing
- 4 time points for each of these two factors identified

### Some of the Challenges Addressed

OBBR Office of Biorepositories and Biospecimen Research

- Development of biospecimen lifecycle Common Data Elements (CDEs) and data enty system
- Operational schemes for labeling and randomization
- Experimental design and execution

Office of Biorepositories and Biospecimen Research

## Developing Common Data Elements for Fixation and Processing Parameters



### **OpenClinica utilized for Data Collection**

Barcode reader functionality added – records a timestamp when reading barcode



OBBR Office of Biorepositories and Biospecimen Research

### • Van Andel Research Institute (VARI)

- caHUB contracted Comprehensive Biospecimen Resource
- Prepare ID's, labels, and randomization tool ("Experimental Key") for BRN
- Perform basic molecular analysis for BRN
- Manage shipment and storage of BRN biospecimens

### IDs and labels:

- BRN case IDs: umbrella ID for all biospecimens collected at one event (one donor on one day)
- BRN biospecimen IDs: all derivative biospecimens from that case
- Random numbers
- Numbers and barcodes placed on labels



- IDs and labels:
  - In most cases, labels are placed on collection and storage containers at the sites
  - FFPE cassettes are pre-etched with (random number) biospecimen IDs at VARI



- Laminated Grids
  - To aid in dissecting tissue into aliquots
- Experimental Keys:
  - To randomly assign aliquots to a particular experimental protocol

Scannable Key ID	
Experimental Protocol	Grid Location
A	9
В	13
С	5
D	10
E	2
F	7
G	16
Н	1
1	11
J	4
К	8
L	14
M	3
N	12
0	6
Р	15



# How ID's, labels, Experimental Keys work with OpenClinica



- Technician scans each biospecimen label into the OpenClinica system
- A date and time stamp populates the appropriate OpenClinica data entry field
- The layout of the Experimental Key is aligned with the OpenClinica data entry form

#### View Section Data Entry for BRN Tissue Processing Worksheet4 Original

bid fissue frocessing worksiteeer original	Discrepancy Notes:	New, Updated, Resolution Proposed, Closed, Not Applicable		A	)			
Study Subject ID:	Person ID:			/-	Y)	012	***	
Study/Site:	Age At				5		7	
Event: 0	Enrollment:	Save			T			
Interviewer Name: *	Interview Date: *	JCX.		P DT				
Press the little flag icon beside an input to enter discre	epancy notes, please note th	nat you can only save the notes if CRF da	ata entry has already started.	-05				
Exit								
Tissue(0/71)  Select to Jump	•							
Title: Tissue Processing Worksheet								
						_		
caHUB/BRN Case ID (OC Study Subject ID)	caHUB/BRN Case ID     B55 Tissue Bank Participant ID     BRN       (OC Study Subject ID)     (OC Secondary ID)     fr		BRN parent sample barcode ID from which required study samples were derived	Experimental Key	Experimental Key Barcode ID		Date and time Experimental Key Barcode ID was scanned	
CA-12345–BRN			12345678-BRN	► KE -123456	-BRN	2011-	09-28T12:02:0	
	L							
Frozen Tissue Sample Information (Bottom 5	Section of Tumor Block)		Γ	Scannable Key ID			1	
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block)	Section of Tumor Block)		[	Scannable Key ID Experimental Prot A	ocol Grid	Location		
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor	Section of Tumor Block)		[	Scannable Key ID Experimental Prot A B	ocol Grid	Location 9 13		
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was	Section of Tumor Block)		[	Scannable Key ID Experimental Prot A B C	ocol Grid	Location 9 13 5		
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen	Section of Tumor Block)		[	Scannable Key ID Experimental Prot A B C D	ocol Grid	Location 9 13 5 10		
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen Planned Delay to Fixation 0-30 Minutes Expe	Section of Tumor Block)	nd Top FFPE Section Part I		Scannable Key ID Experimental Prot A B C D F	ocol Grid	Location 9 13 5 10 2		
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen Planned Delay to Fixation 0-30 Minutes Expe Experimental Protocol A-D And Top FFPE Section	Section of Tumor Block) erimental Protocol A-D an Planned delay to fixation time	nd Top FFPE Section Part I Scanned ID of cassette: Record first scan	Time that cassette was first scanned	Scannable Key ID Experimental Prot A B C D E F Scanned ID of cassette: Record time placed in fixative H	ocol Grid	Location 9 13 5 10 2 assette to flegive red) 1	OPTIONAL: Calculated actual delay to fixation time	
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen Planned Delay to Fixation 0-30 Minutes Expe Experimental Protocol A-D And Top FFPE Section	Erimental Protocol A-D ar Planned delay to fixation time	nd Top FFPE Section Part I Scanned ID of cassette: Record first scan	Time that cassette was first scanned	Scannable Key ID Experimental Prot A B C D E F Scanned ID of cassette: Record time placed in fixative H	ocol Grid	Location 9 13 5 10 2 7 assette to 16 Give ed) 1 8 = 17:45:0	OPTIONAL: Calculated actual delay to fixation time	
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen Planned Delay to Fixation 0-30 Minutes Expe Experimental Protocol A-D And Top FFPE Section	Erimental Protocol A-D an Planned delay to fixation time 0-30 minutes •	nd Top FFPE Section Part I Scanned ID of cassette: Record first scan 12345678-BRN	Time that cassette was first scanned	Scannable Key ID Experimental Prot A B C D E F scanned ID of cassette: Record time placed in fixative H	ocol Grid	Location 9 13 5 10 2 assette to 16 Give ed) 1 8 # 17:45:0 4	OPTIONAL: Calculated actual delay to fixation time	
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen Planned Delay to Fixation 0-30 Minutes Expe Experimental Protocol A-D And Top FFPE Section	Erimental Protocol A-D ar Planned delay to fixation time	nd Top FFPE Section Part I Scanned ID of cassette: Record first scan 12345678-BRN	Time that cassette was first scanned	Scannable Key ID Experimental Prot A B C D E F Scanned ID of cassette: Record time placed in fixative H	ocol Grid	Location 9 13 5 10 2 assette to fictione ed) 1 8717:45:0 4 8	OPTIONAL: Calculated actual delay to fixation time	
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen Planned Delay to Fixation 0-30 Minutes Expe Experimental Protocol A-D And Top FFPE Section Protocol A' <6 hours in fixative Select	Erimental Protocol A-D an Planned delay to fixation time 0.30 minutes • 0.30 minutes • 0.30 minutes •	nd Top FFPE Section Part I Scanned ID of cassette: Record first scan 12345678-BRN	Time that cassette was first scanned	Scannable Key ID Experimental Prot A B C D E F Scanned ID of cassette: Record time placed in fixative H	ocol Grid	Location 9 13 5 10 2 assette to fleGive ed) 1 8 7 17:45:0 4 8 14 2	OPTIONAL: Calculated actual delay to fixation time	
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen Planned Delay to Fixation 0-30 Minutes Expe Experimental Protocol A-D And Top FFPE Section Protocol A: <6 hours in fixative Select	erimental Protocol A-D ar Planned delay to fixation time 0-30 minutes • 0-30 minutes • 0-30 minutes • 0-30 minutes •	nd Top FFPE Section Part I Scanned ID of cassette: Record first scan 12345678-BRN	Time that cassette was first scanned	Scannable Key ID Experimental Prot A B C D E F Scanned ID of cassette: Record time placed in fixative H 2345678-BRN J K L M	ocol Grid	Location 9 13 5 10 2 7 assette to 16 ive edi 1 8 14 3 12	OPTIONAL: Calculated actual delay to fixation time       D       I       I       I       I       I       I       I       I       I       I       I       I       I       I       I	
Frozen Tissue Sample Information (Bottom S Scanned ID of frozen tumor tissue cryosette (bottom section of tumor block) Time that tumor tissue sample was frozen in liquid nitrogen Planned Delay to Fixation 0-30 Minutes Expe Experimental Protocol A-D And Top FFPE Section Protocol A: <6 hours in fixativ/ Select	Erimental Protocol A-D ar Planned delay to fixation time 0-30 minutes • 0-30 minutes • 0-30 minutes • 0-30 minutes • 0-30 minutes •	nd Top FFPE Section Part I  Scanned ID of cassette: Record first scan  12345678-BRN	Time that cassette was first scanned       I         V       2011-09-28T12:35	Scannable Key ID Experimental Prot A B C D E F Scanned ID of cassette: Record time placed in fixative H 2345678-BRN J K L M N	ocol Grid	Location 9 13 5 10 2 7 7 3 sette to fefore ed) 1 8 7 7 :45 0 4 8 14 3 12 6	OPTIONAL: Calculated actual delay to fixation time       D       I	

### **Tissue Processor**

### OBBR Office of Biorepositories and Biospecimen Research

• Leica Peloris

### Can perform multiple runs in parallel

- So we can do a 6 pm and a midnight run on the same machine
- One machine to maintain and QC

#### Uses standard reagents

- No need for Leica-branded consumables
- Can do isopropanol and xylene-free runs later in project, if desired
- Reagents for different runs are pulled from the same "bucket" of reagent
  - One set of stepwise reagents to maintain and QC

Step	Reagent type	Reagent group	Time (min)	Temp (°C)	P/V	Stirrer	Drip time (s)
1	Formalin	Fixatives	20	45 )??	Ambient	Medium	10
2	Ethanol	Dehydrants	20	45	Ambient	Medium	10
3 4 5 6 7 8	ER recomment tissue process processor contesting. If such responsible for been processor (Temperatures stained.)	dations, 200 sor solutions ntains breast h heated pro- or validation ed conventions above 37C	8: It is str s, excludin tissue for cessors a of ER rest nally with are not re	ongly rec ng paraffin r potentia re used, t ults again out exces commend	ommende ns should I ER and o hen the la st paralle ss heat. ded for tis	ed that non l exceed 37 other biom aboratory o l samples t sue that w	e of the C if the arker director is that have
9 <sup>L</sup>	хуюне	clearers	30	40	Ampient	meaium	10
10	Xylene	Clearers	60	45	Ambient	Medium	10
11	Paraffin wax	Wax	40	65	Vacuum	Medium	10
12	Paraffin wax	Wax	40	65	Vacuum	Medium	10
13	Paraffin wax	Wax	60	65	Vacuum	Medium	10
Proce	ssing time		8:08:00				



Components need to collect the first tissues: Experiment 1

- ✓ IRB Approvals: UNM and Vanderbilt
- Project equipment in place (including tissue processor)
- BRN data collection tool (OpenClinica) tested and ready to go
- Biospecimen ID's, labels, randomization tools ready to go
- SOPs and other deployment and training materials complete
- ✓ Internal caHUB approvals processes complete
- Training conducted late October/early November 2011
- Currently collecting for Experiment 1 at both sites

## Thanks to...

### $\overline{\mathsf{OBBR}}$ Office of Biorepositories and Biospecimen Research

- Therese Bocklage
- Carolyn Compton
- Kelly Engel
- Erin Gover
- Howard Greenman
- Kelly Higgins
- Andrea Kelly
- Nicole Lockhart
- Cathleen Martinez
- Marlena Martinez
- Neil Mucci
- Angela Notah-Chavez

- Nancy Roche
- Anna Smith
- David Tabor
- Jake Vargas
- Jim Vaught
- Kay Washington
- Carol Weil
- Kerry Wiles
- Gail Wiseman
- Jennifer Hunt and the BRN Scientific Steering Committee

# OBBR Office of Biorepositories and Biospecimen Research



# BRN Tissue Acquisition Variables Project

Helen M. Moore, PhD January 24, 2012

