



# **Rapid Methods for the Assessment of Tissue Quality**

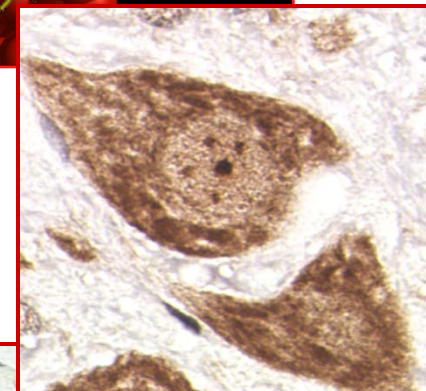
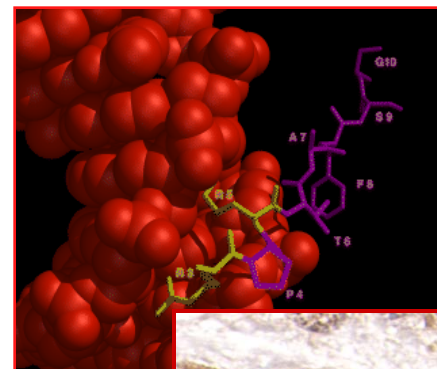
Charles Saller, Ph.D.

Analytical Biological Services Inc.

President & CEO

# Biospecimen Services

- ✓ Tissues, blood, and fluids
- ✓ RNA and DNA
- ✓ Protein Extracts
- ✓ Subcellular Fractions
- ✓ Histology
- ✓ Immunohistochemistry
- ✓ In situ hybridization
- ✓ Tumor Cell Lines
- ✓ Primary Tumor Screening
- ✓ Tissue Quality Analysis





# Collaborators

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## **Tissue Collections**

George Sandusky, DVM, PhD (IUSCC)  
Colleen Mitchell (IUSCC)  
Katherine Carr, MPH (ABS Indiana)

## **Quality Assurance**

John Yergler, MS (ABS)

## **Biochemical Assay R&D (ABS)**

Linda Carlini  
David Satinover, PhD  
Christopher Orr, PhD.

## **IHC/ISH**

Slidomics, LLC

## **Reverse Phase Protein Arrays (Center for Applied Proteomics and Molecular Medicine, GMU)**

Lance Liotta, PhD, MD  
Chip Petricoin, PhD  
Virginia Espina, PhD

## **Computational Analysis**

Tudor Oprea, MD, PhD,(UNM, Sunset Molecular, LLC)  
Jake Chen, PhD (Indiana University)

# Presentation Outline

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- **What:** Project Goals
- **Why:** Sample Quality Challenges
  - Heterogeneity issues with human biospecimens
    - Population heterogeneity
    - Sample heterogeneity
  - Collection and processing challenges
- **How:** Experimental Approach

# Project Goals

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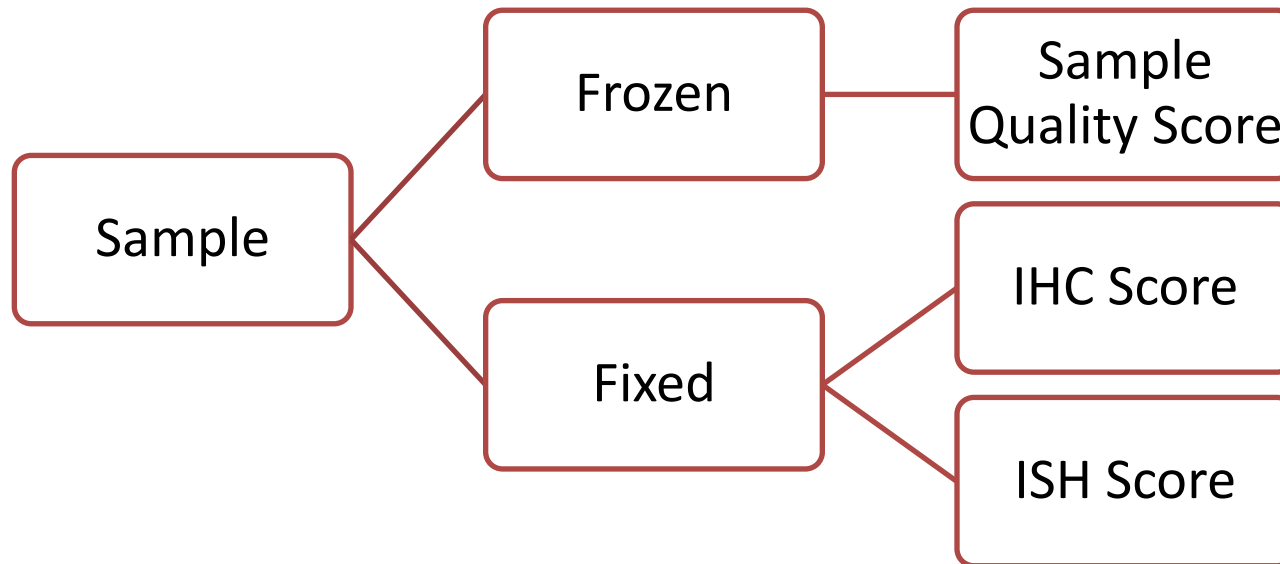
**Develop Methods to Quickly Assess the Quality of Frozen and Fixed Tissue Samples**

**→ Sample Quality Scores**

- Fast
- Inexpensive
- Use small amounts of tissue
- Easy to Use (mix & read, multiplexed)
- Highly predictive of “Fit for Purpose”

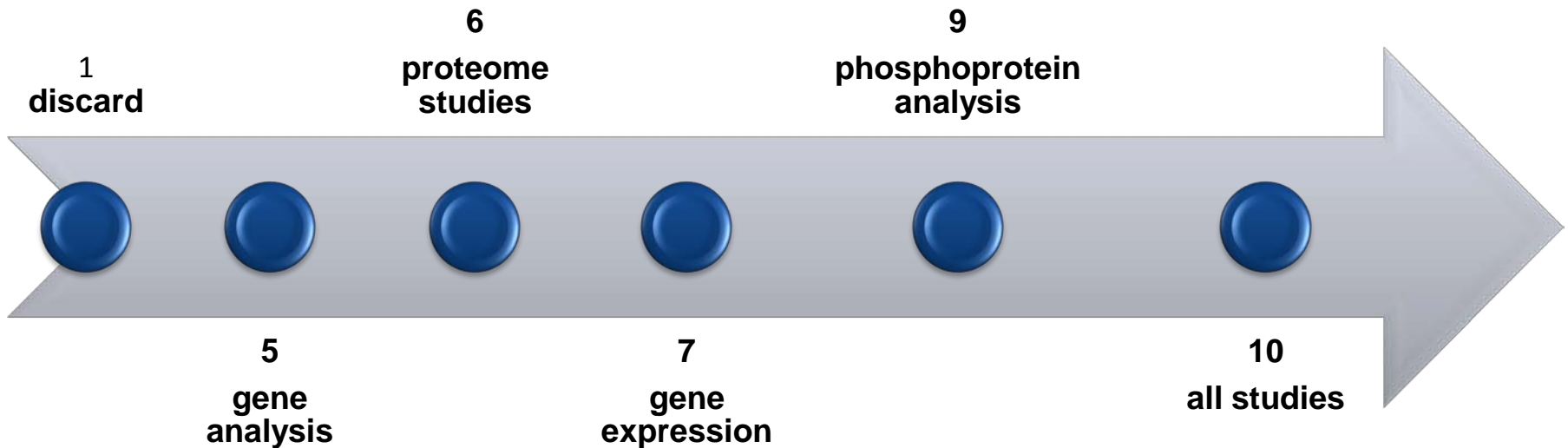
# Quality Scores

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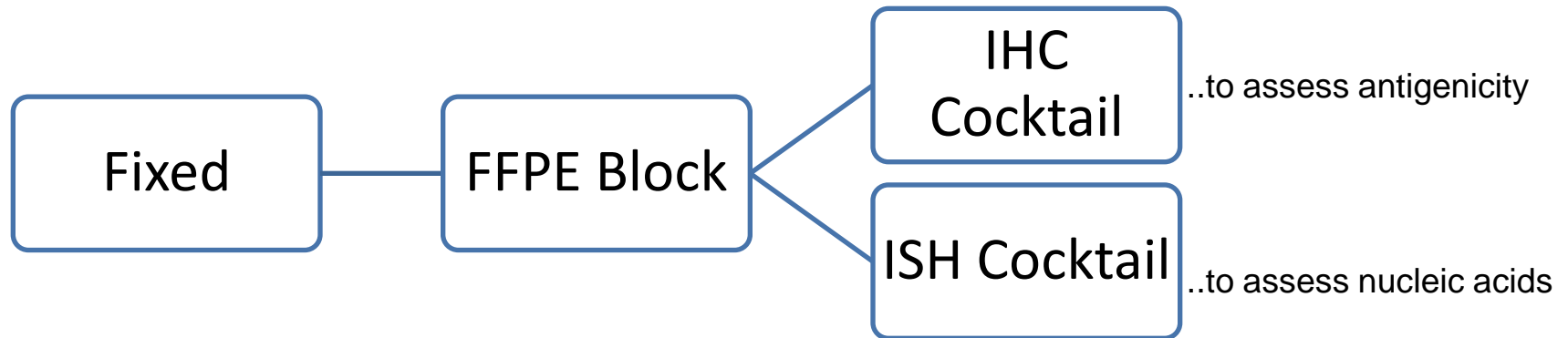
# Sample Quality Score (SQS)

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hypothetical example

# Histological Scoring



IHC cocktail = mixture of antibodies to label many cell types and proteins (actins, keratins, vimentin, inflammatory cells and proliferating cells)

ISH cocktail =  $\beta$ -actin and GAP-DH, RNA probes

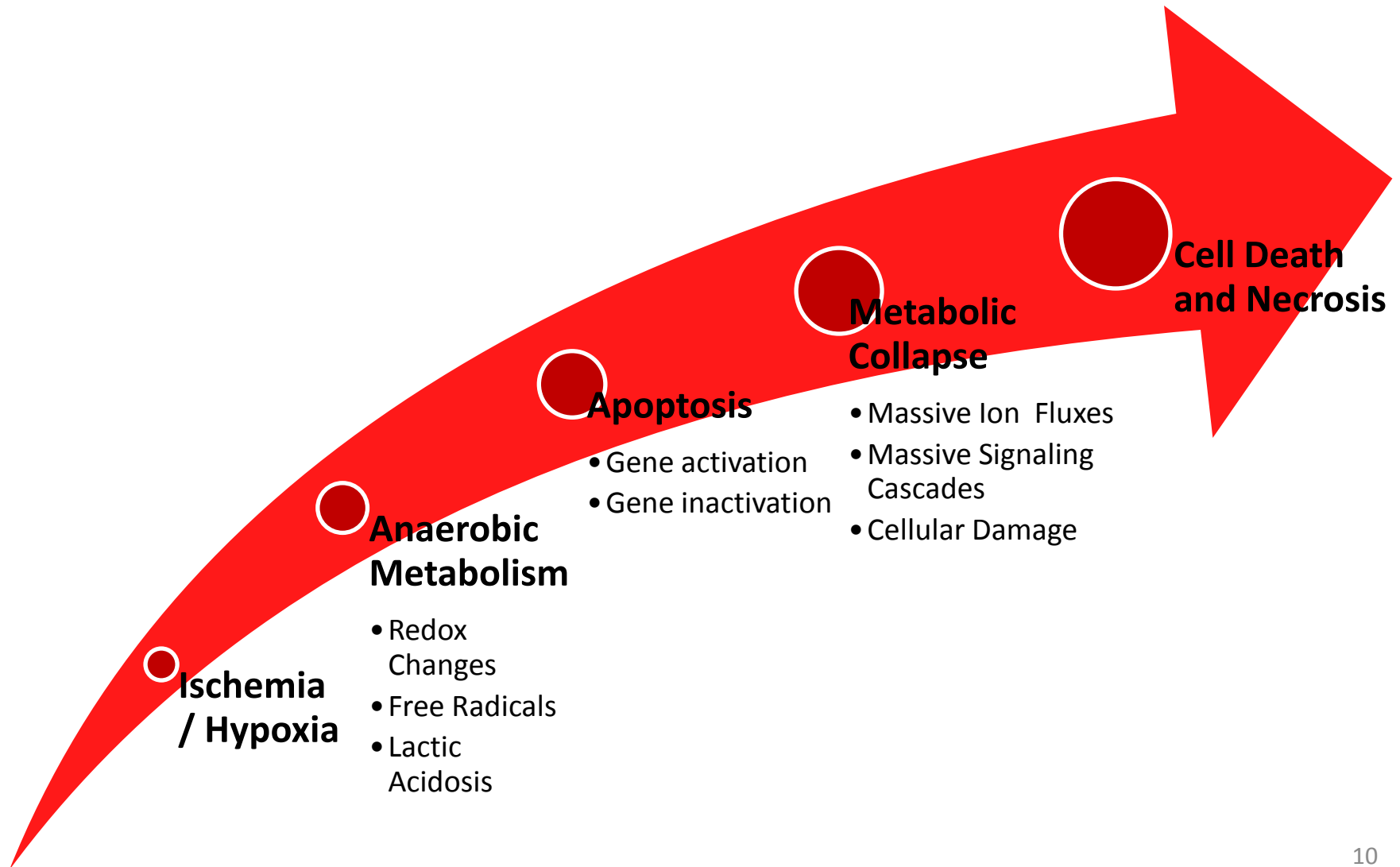


# What is Tissue Quality?

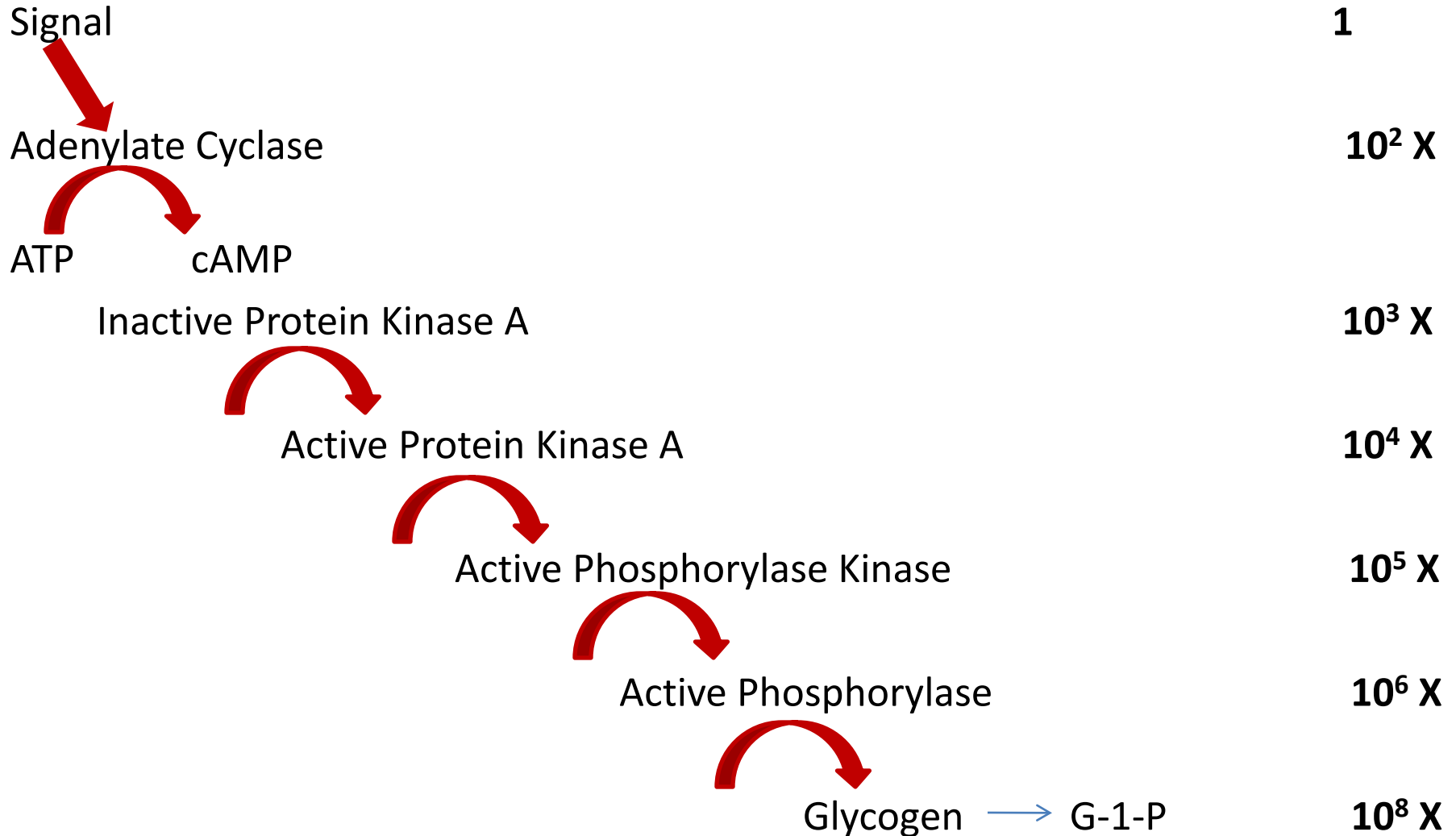
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- How do you know if a sample is “Good or Bad”?
- What does “Fit for Purpose” mean? Can a tissue be OK for one method (IHC) and not for another method (RNA) at the same time?
- How do you know if the signaling pathway (gene, etc.) changes you are measuring are a consequence of the disease or if it is related to hypoxia and/or ischemia, medications, anesthesia, etc.?

# Post-Ischemia / Hypoxia Deterioration



# Phosphorylation Cascades



# Transcription Factors Induced by cAMP or Hypoxia

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- Hypoxia Inducible Factor (HIF-1)
- NF- $\kappa$ B
- cAMP Response Binding Proteins (CERB), including Activating Transcription Factor (ATF-1)
- cAMP Response Element Modulator (CREM)
- Steriodogenic Factor (SF-1)

# Solutions

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- Standardization & Annotation
- Importance of Adjacent Tissues
- Fast, Easy, Reliable Assays
- Multivariate Analysis



# Standardization & Annotation

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## Collection Variables

Clamp time  
Time of excision  
Time of freezing  
Fixation time after excision  
Length of fixation period  
Approximate sample size  
Sample Temperature  
Number of sample aliquots

## Clinical Variables

Anesthesia  
Radiation therapy  
Chemotherapy  
Pre-surgical medications  
Histology  
Diagnosis  
Tumor Staging  
Race, Ethnicity

# Importance of Adjacent Tissues

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- Not “normal”, but an important internal control
- Exposed to same pre-surgical variables as tumor
- Exposed to same surgical variables (anesthesia, etc.)
- Exposed to similar processing post-excision
- Exposed to same analytical procedures

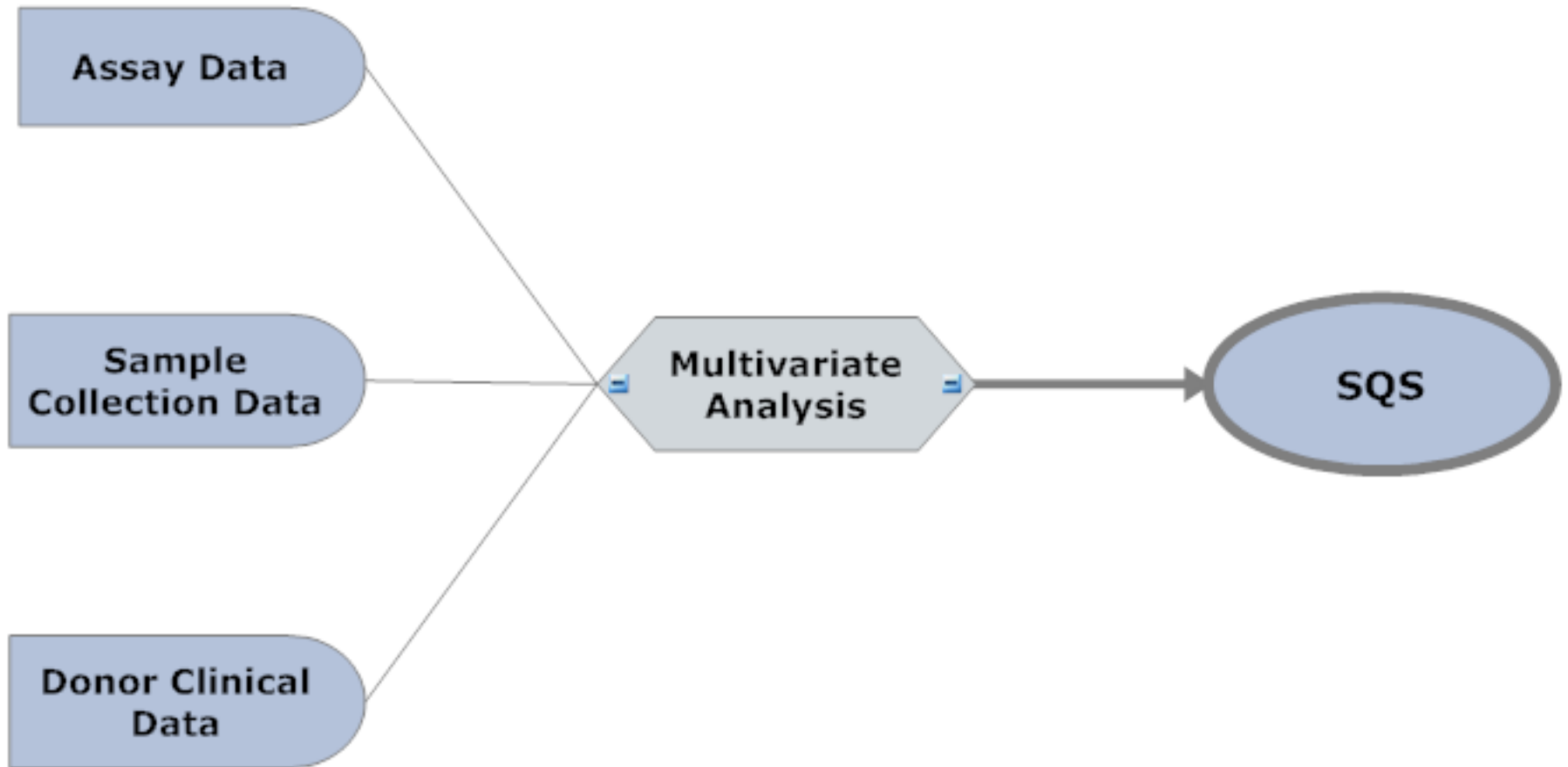
# Sample Criteria

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- > 70% viable tumor nuclei
- <20% of the viable cells should be normal stromal, inflammatory or immune cells
- < 20% Necrosis
- < 30 min. Tissue Preservation Interval  
(i.e., warm + cold ischemia time), unless otherwise required for validation studies

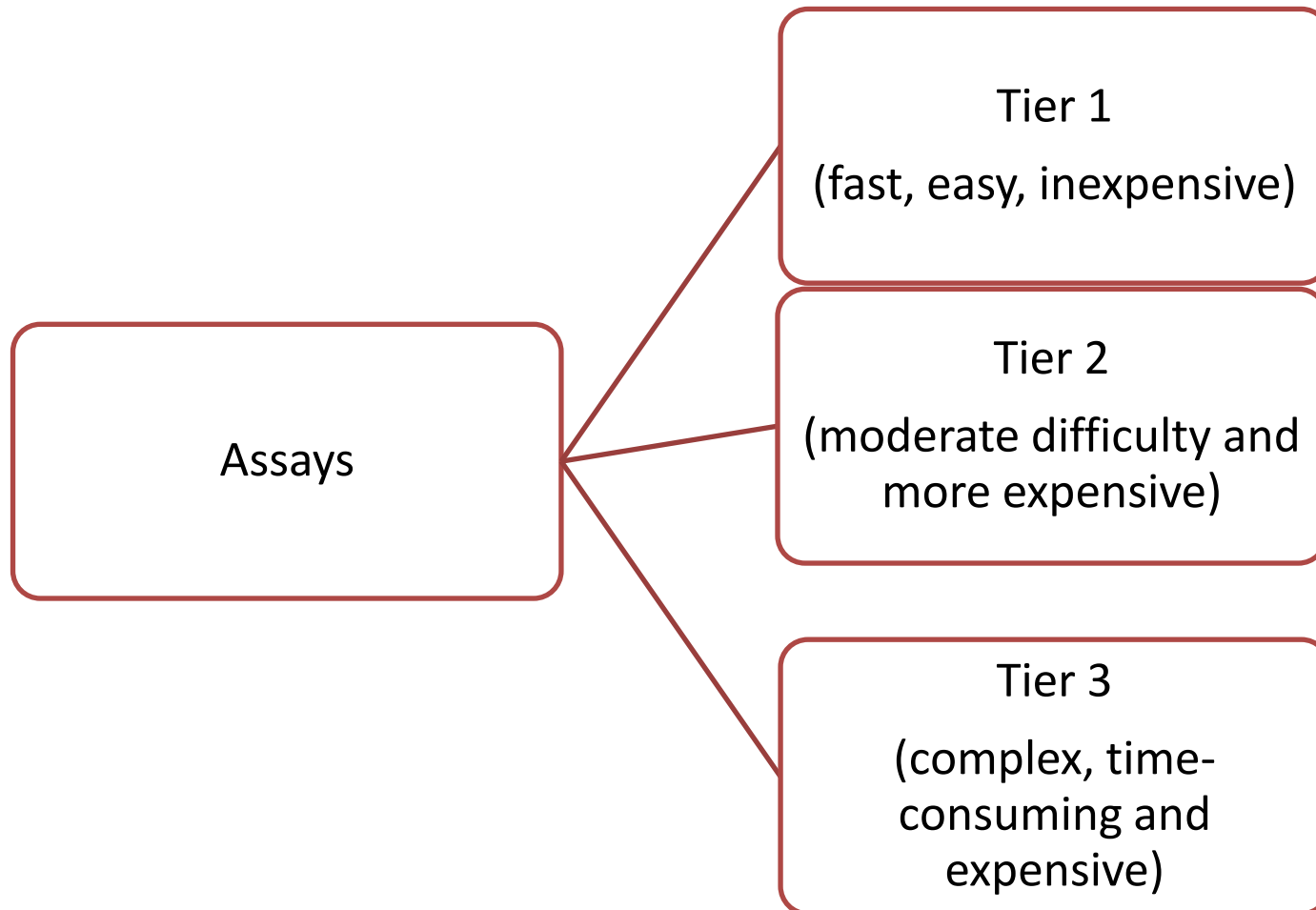


# SQS Paradigm



# Tissue Quality Assays

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# Tier 1 Assay Criteria

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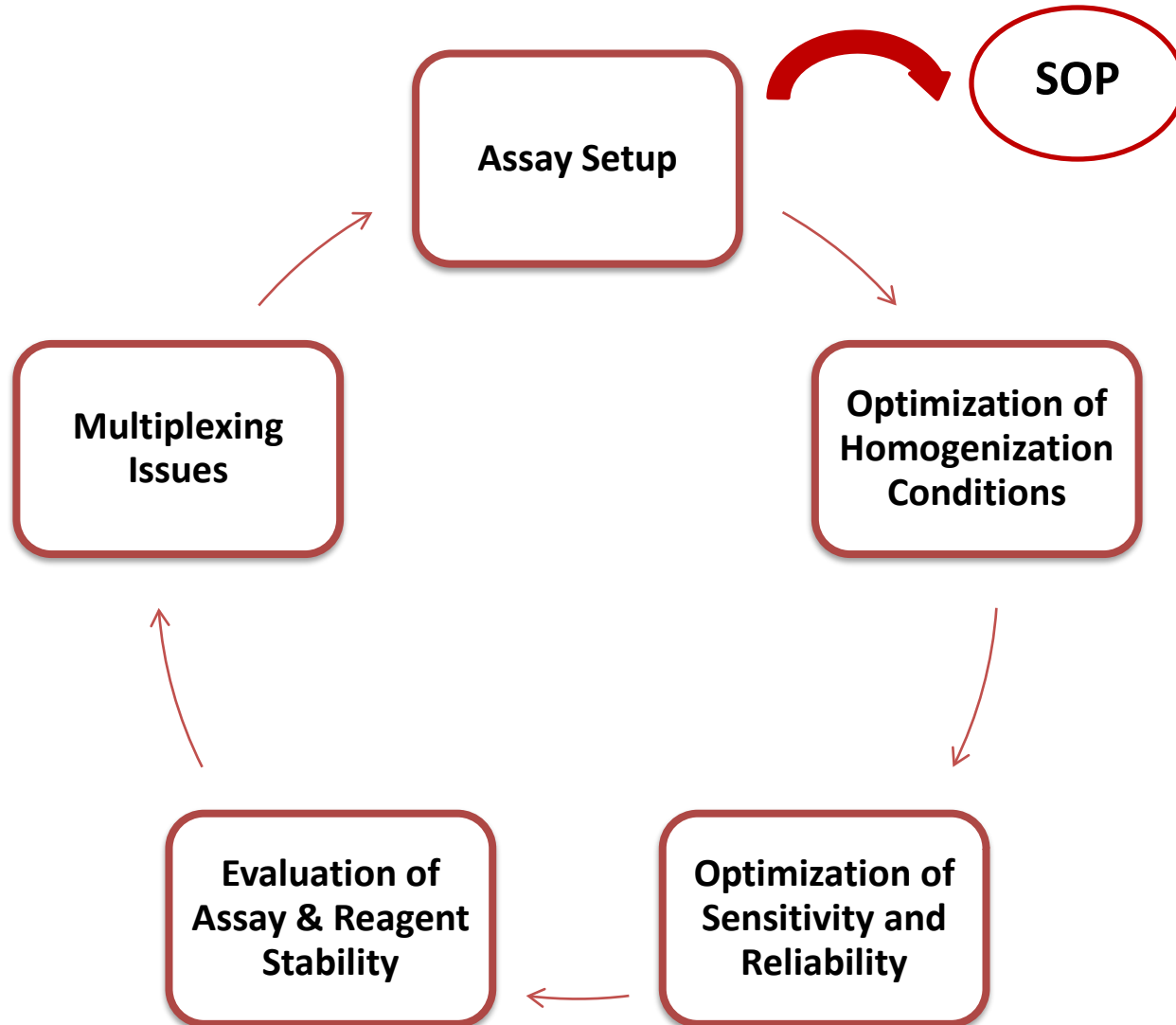
- Low Cost
- Mix and Read
- Reagents stable
- No rigorous timing requirements
- Large linear range as function of protein conc.
- Sensitivity (requires < 1 mg tissue, including replicates)
- 384 well compatible
- Multiplexed
- Accurate and Reliable
- **Predictive** ( $\Delta$  as function of Tissue Preservation Interval)

# Tier 1 Assays

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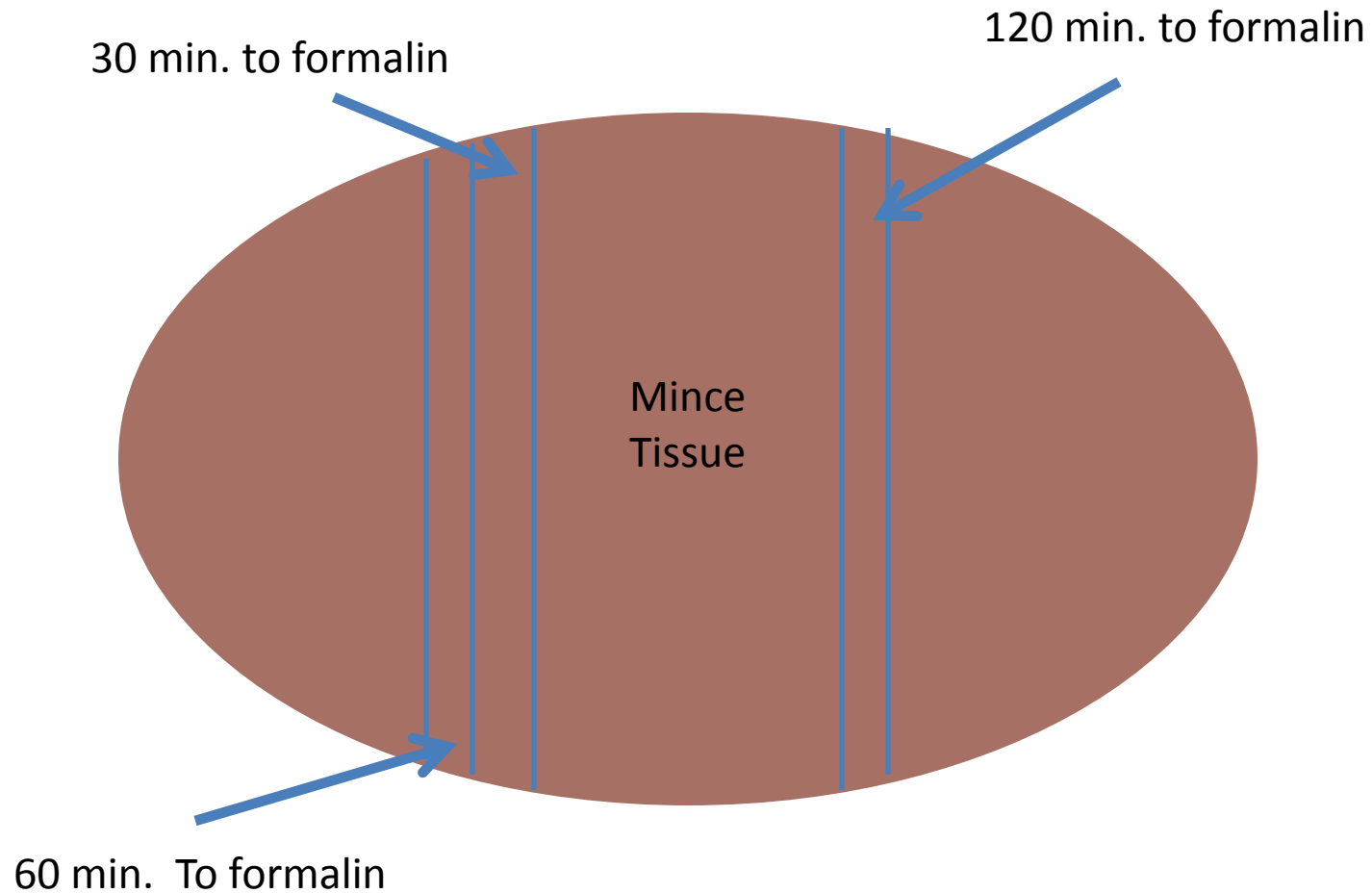
- pH (microelectrode or fluorophore)
- Lactate
- Oxidation-Reduction state (microelectrode)
- Redox Capacity
- Thiol Status
- Lipid Peroxidation
- Protein Degradation

# Tier 1 Assay Development Strategy (Iterative Rounds)



# TPI Experiments

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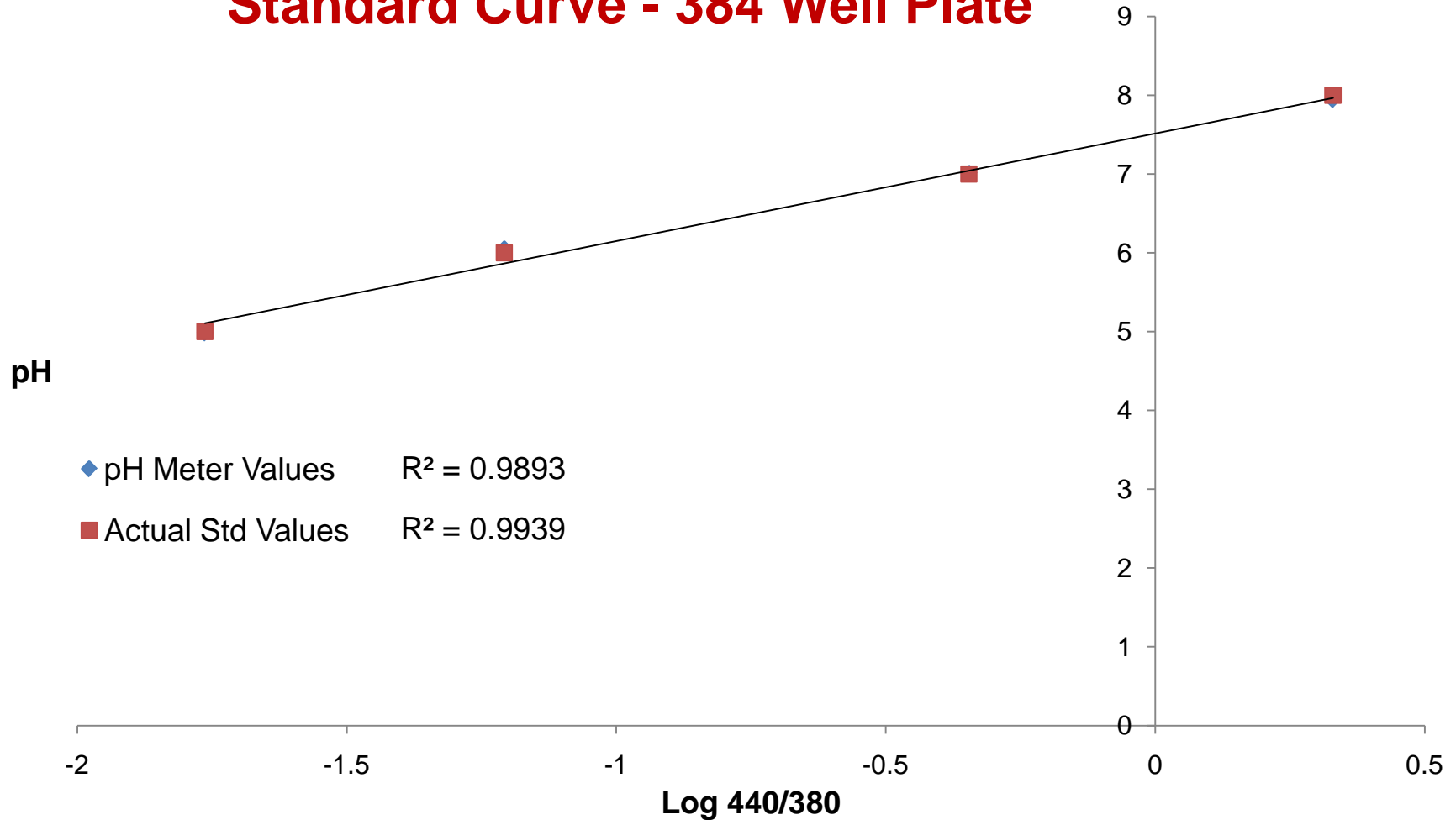






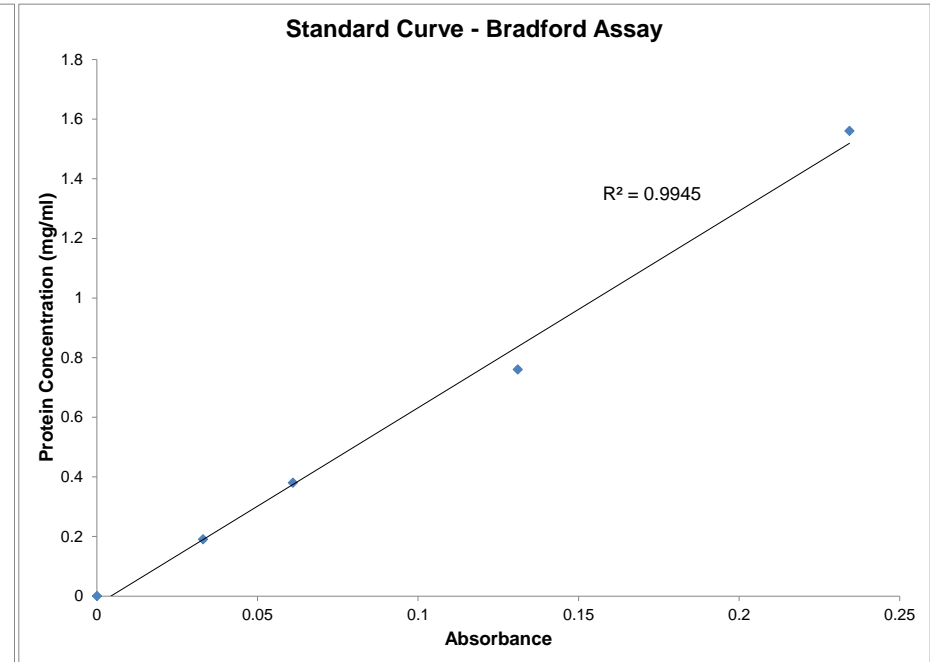
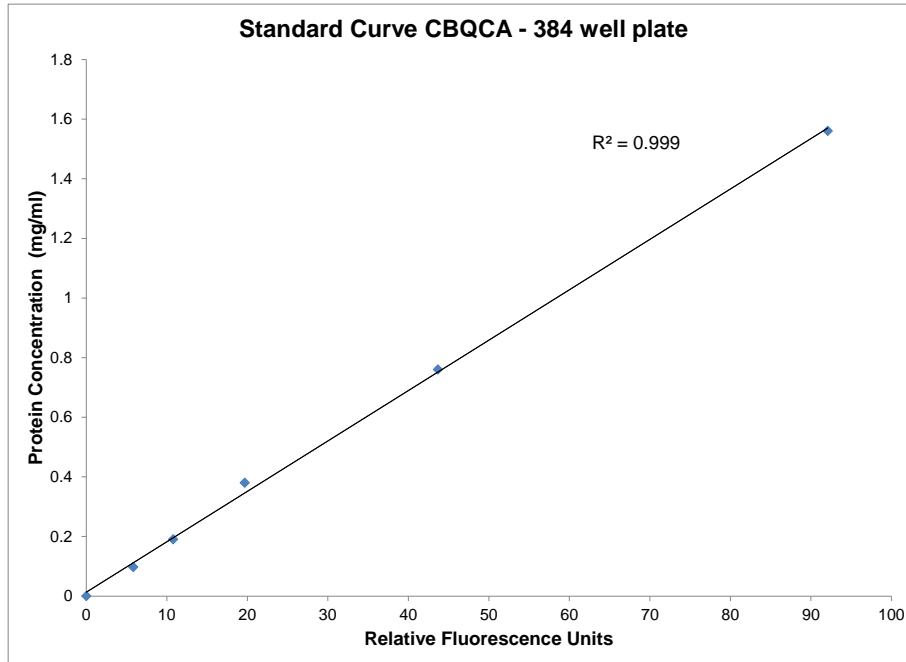
# pH with Microelectrode vs. Fluorophore

## Standard Curve - 384 Well Plate



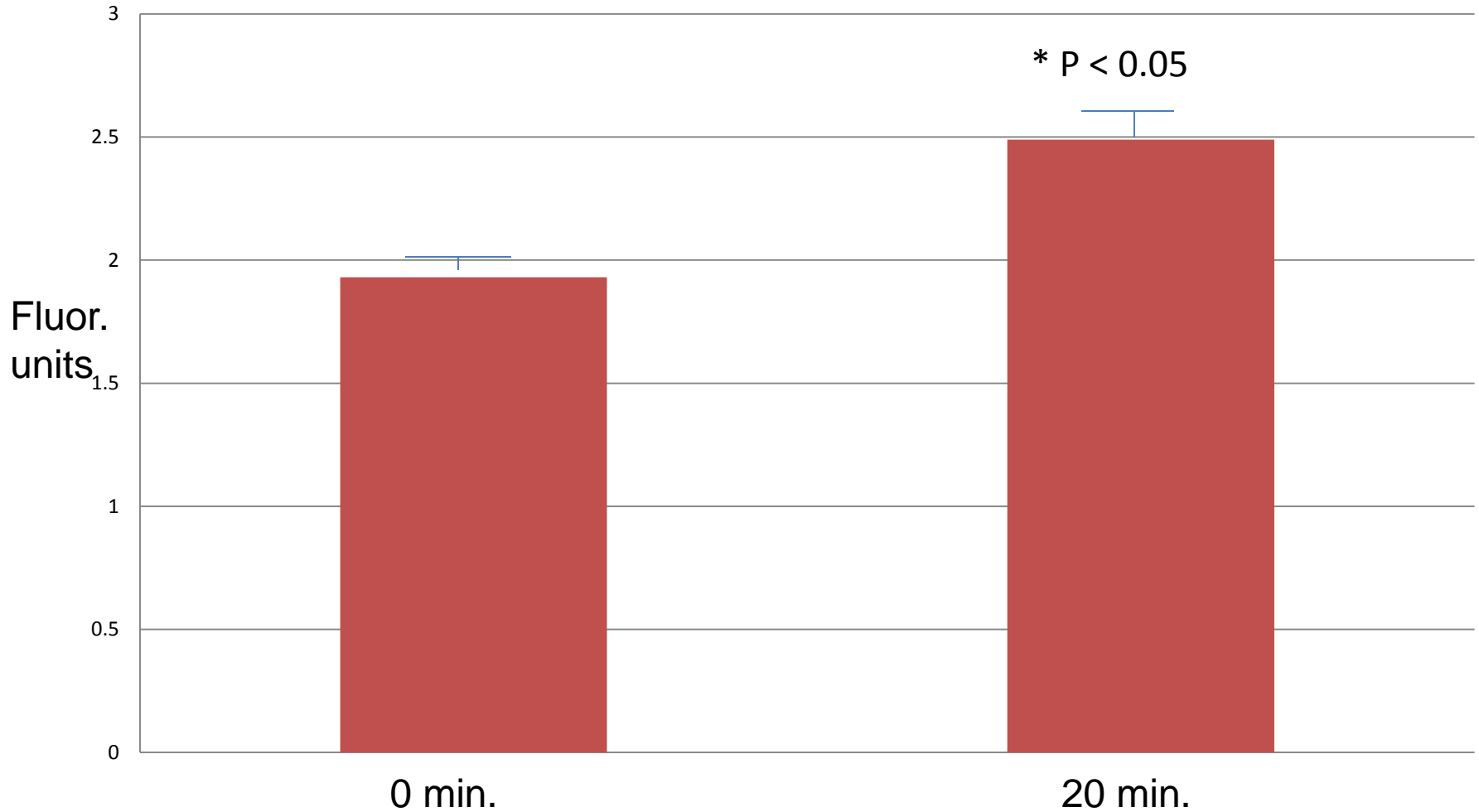


# CBQCA vs. Bradford Assays

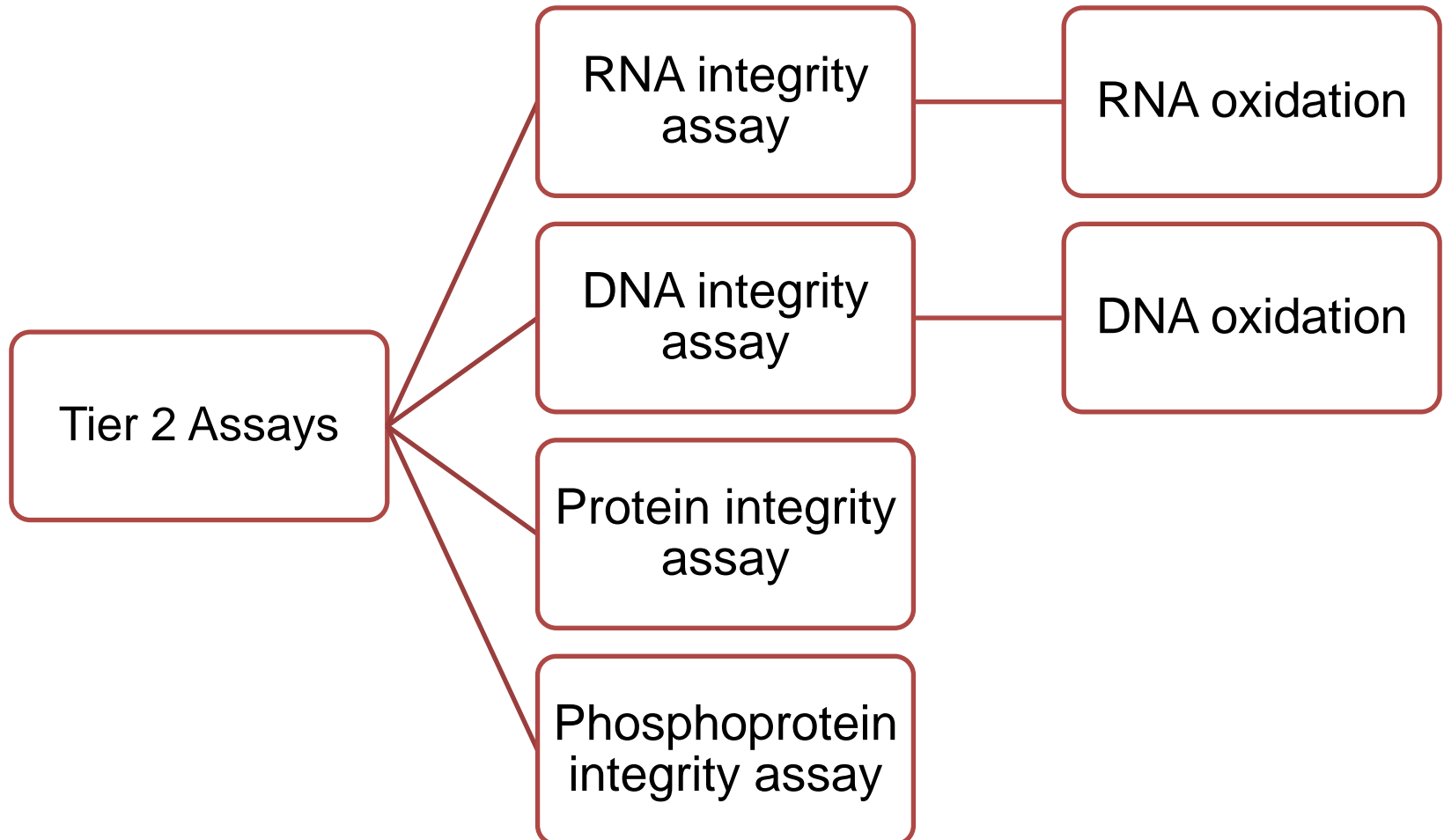


Tissue Type	CBQCA (mg/ml)	Bradford Assay (mg/ml)	CBQCA/Bradford Assay Ratio
Lung	1.706298	0.999288	1.707514
Pancreas	2.102434	1.734567	1.21208
Colon	0.703509	0.338858	2.076116

# Effect of Incubation on CBQCA Assay

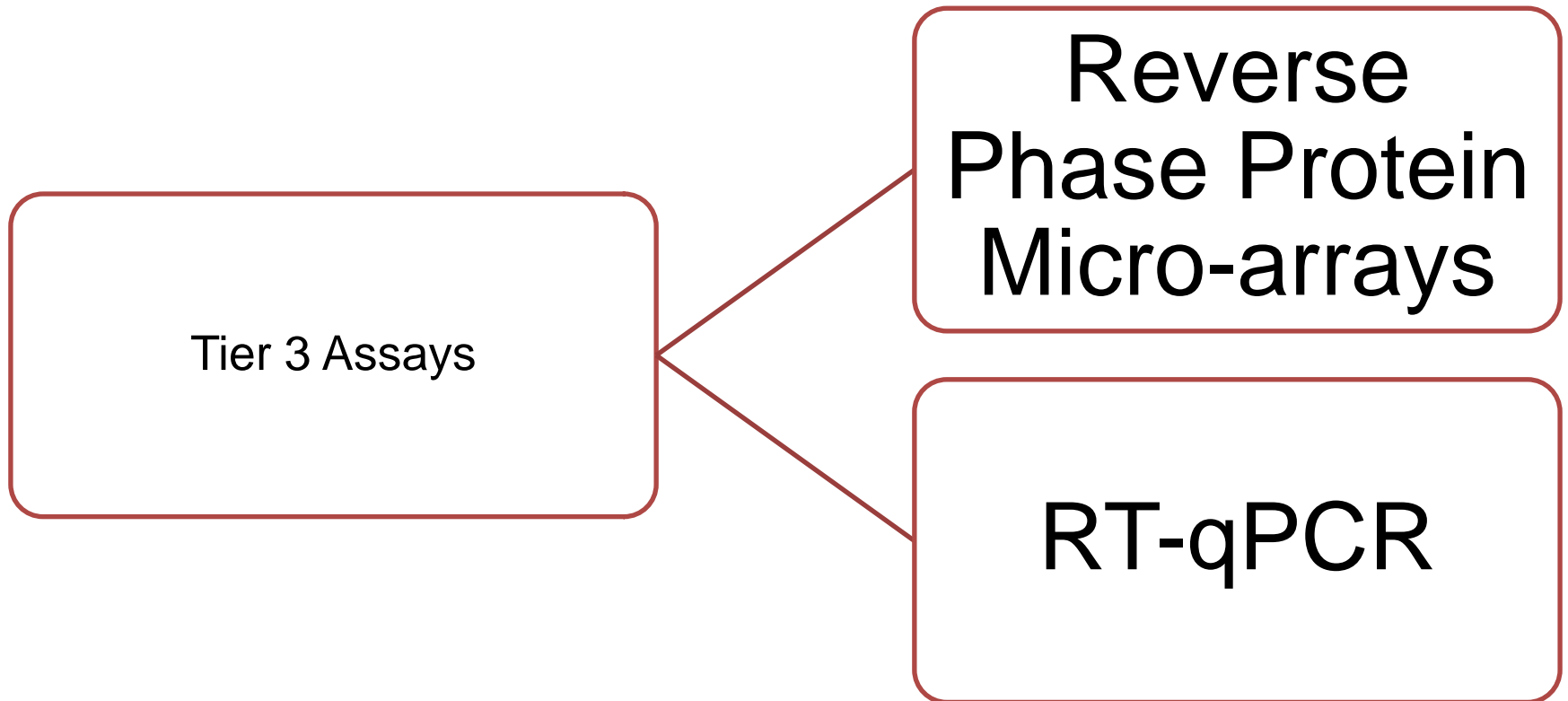


# Tier 2 Assays



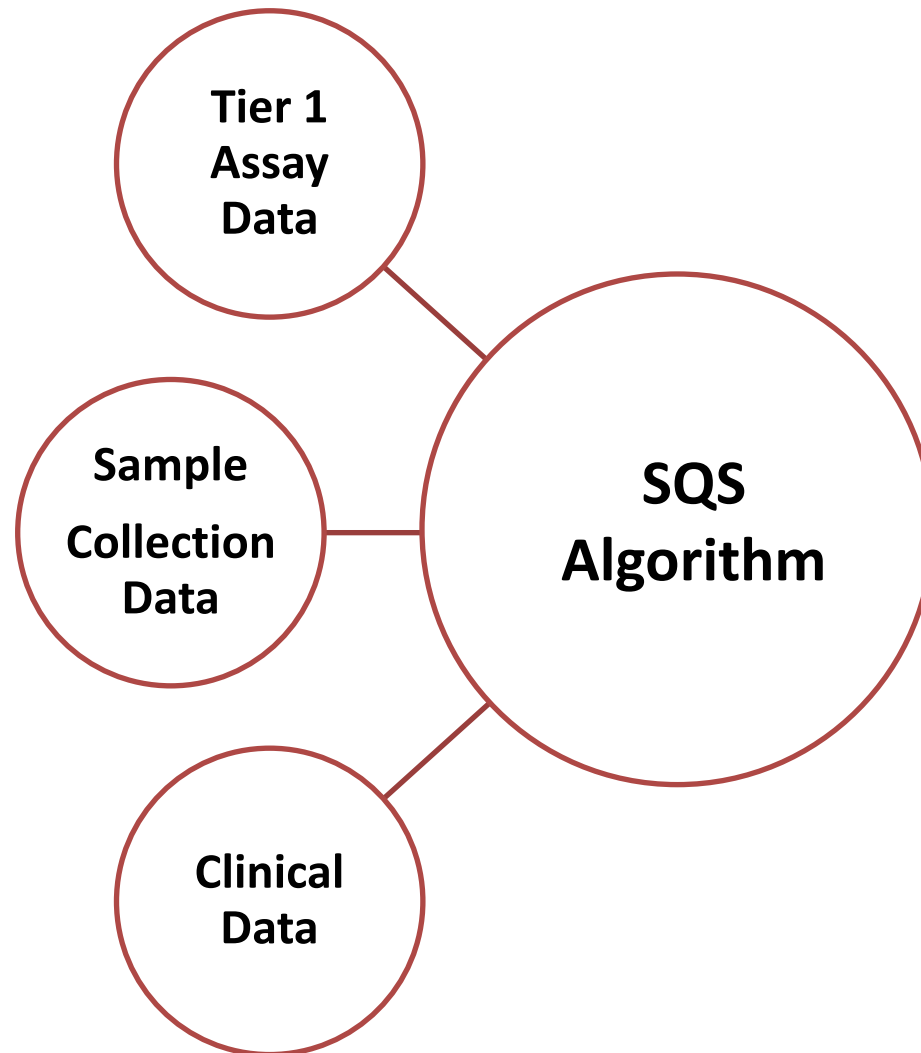
# Tier 3 Assays

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# Creation of SQS Algorithm

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# Data for Multivariate Analysis

## Clinical Data

Specimen Type  
Preparation (Fixed or Frozen)  
Organ of Origin  
Histology Type  
Diagnosis  
Gender  
Age at surgery  
Race  
Ethnicity  
History of Malignancy  
Performance Status  
Tumor Staging (TMN)  
Radiation Therapy  
Chemotherapy  
Anesthesia  
Medications

## Sample Collection Data

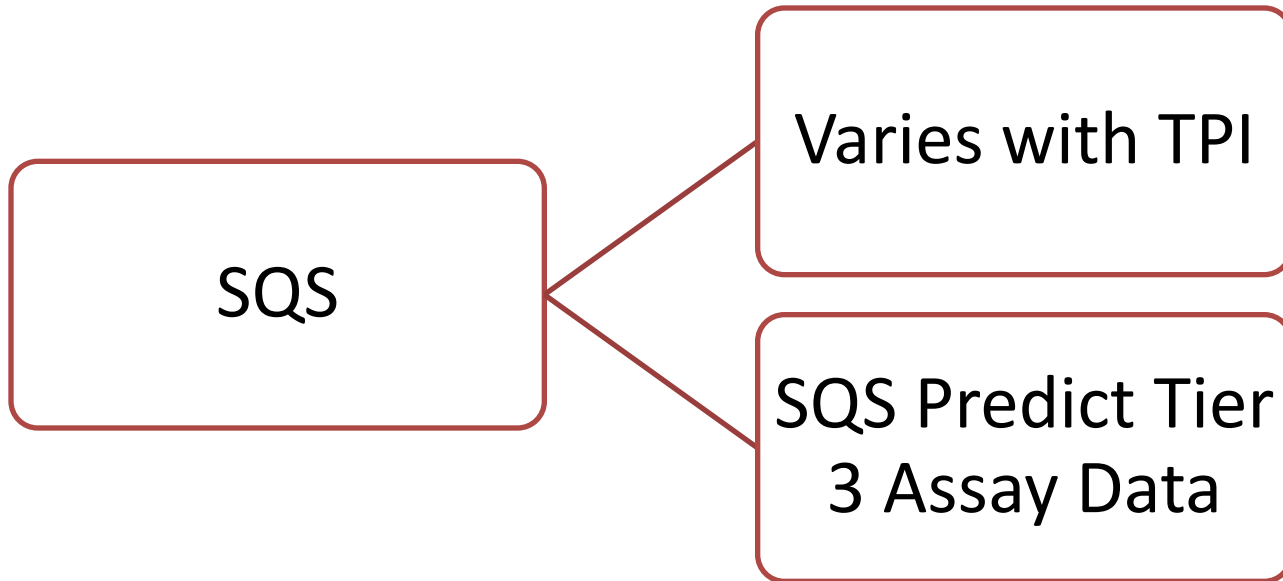
Warm Ischemic Time  
Cold Ischemic Time  
Tissue Preservation Interval  
Sample Weight  
Sample Photography  
Length Fixation  
% Tumor nuclei  
% Inflammation  
% Necrosis

## Analytical Data

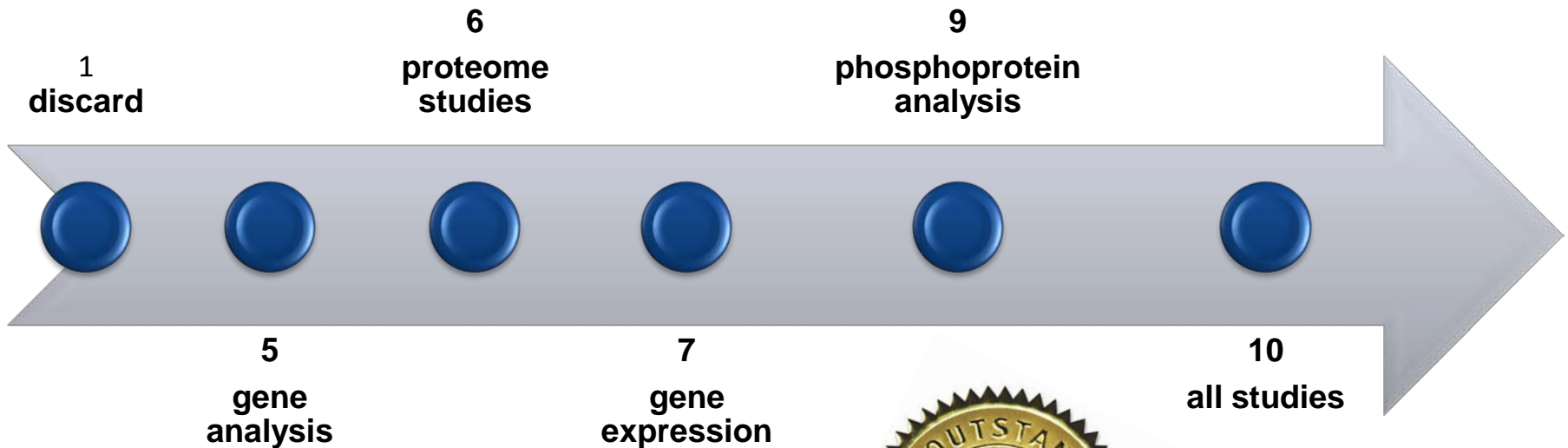
pH  
Lactate  
Oxidation-Reduction state  
Redox Capacity  
Thiol Status  
Lipid Peroxidation  
Protein Degradation  
  
RNA integrity  
DNA integrity  
Phosphorylation Status  
  
RT-qPCR  
Protein Microarrays

# SQS Validation

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# Sample Quality Score (SQS)



hypothetical example

