

# **Research and Development on Human Biospecimen Integrity**

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**BRN Symposium**

**March 29, 2011**

# PROGRAM OVERVIEW



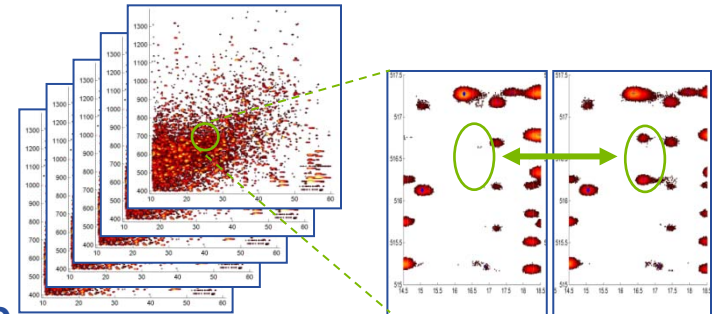
## Overall goal:

- Best practices for proteomics analysis of blood-derived biospecimens
  - Collection, manipulation and storage of samples
    - Guidelines to follow
    - Sample quality assessment assay
    - Tools to facilitate each step

# MS-BASED BIOMARKER DISCOVERY PROCESS

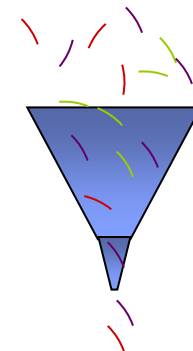
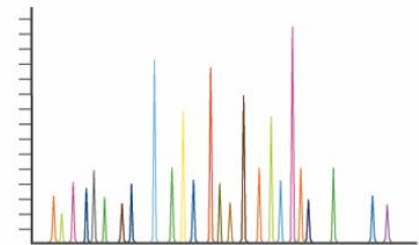
## Biomarker Discovery

- Label-free, gel-free quantitative mass spectrometry
- Non-hypothesis based discovery approach
- Profile 1000's of proteins in 100's samples
- Identify differentially expressed proteins as candidate biomarkers

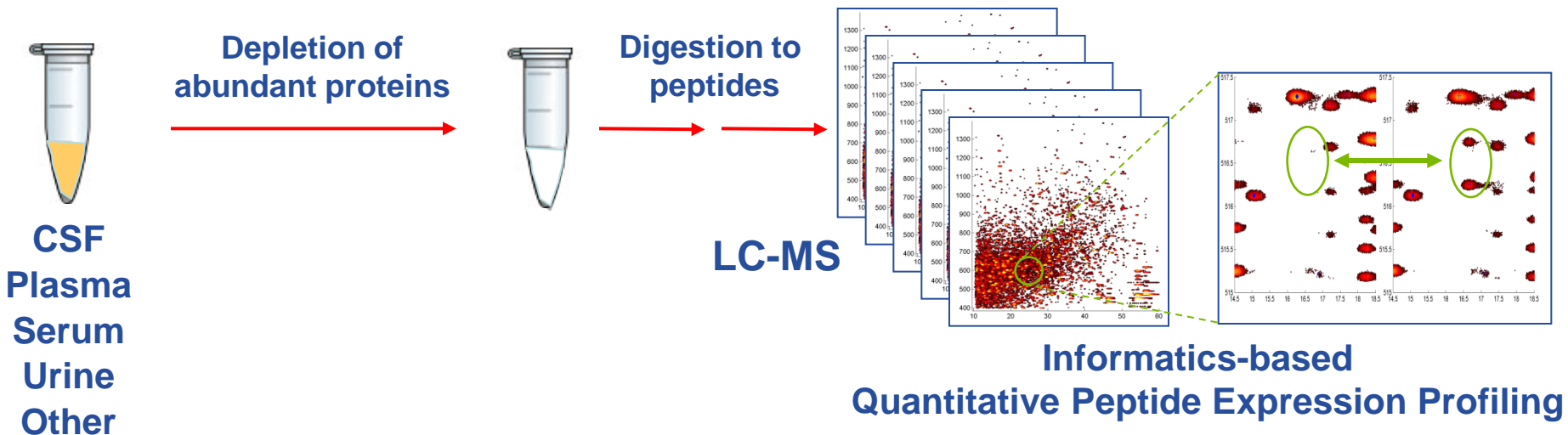


## Multiplexed Assays

- Quantify 1 to 700 proteins in a single “MRM” assay
- Proteins from mass spec, literature, transcriptomics, etc.
- Rapid assay development
- Ab-free or ab enrichment
- Synthetic labeled standards for absolute quantification
- Profile candidate markers in 1,000's samples



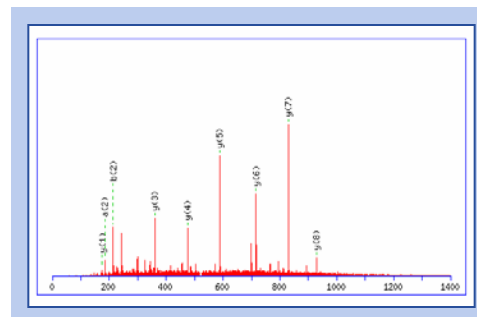
# BIOMARKER DISCOVERY PROCESS: PROTEOMIC EXPRESSION PROFILING



Statistical Analysis

```
>gi|41327732|ref|NP_958439.1|
epidermal growth factor
receptor isoform b [Homo
sapiens]
MRPSGTAGAALLALLAALCPASRALEEKKVCQGTSNKL
TQLGTFEDHFLSLQRMFNCEVVLGNLEITYVQRNYDL
SFLKTIQEVAGYVLIALNIVERIPLNLQIIRGNMYE
NSYALAVLSNYDANKTGLKELPMFNLQEILHGAVRFSN
NPALCNVESIQWRDIVSDFLSNMMDFQNHGSCQKC
DPSCPNGSCWGAGEENCQKLTKTICAQQCSGRCRGKSP
SDCCHNQCAAGCTGPRESDCLVCRKFRDEAT.....
```

Protein Identification



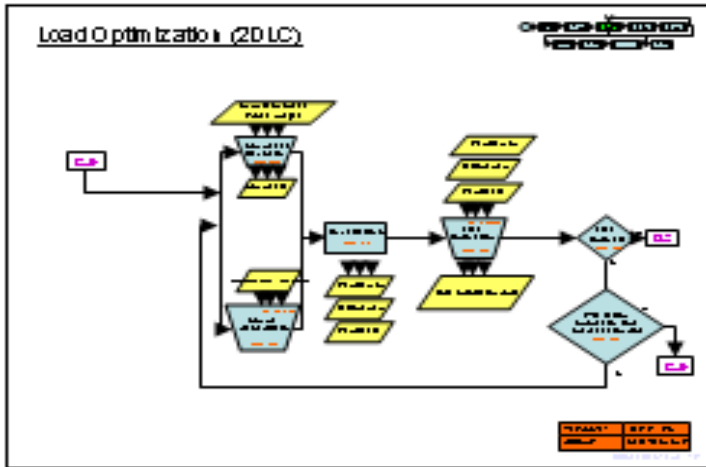
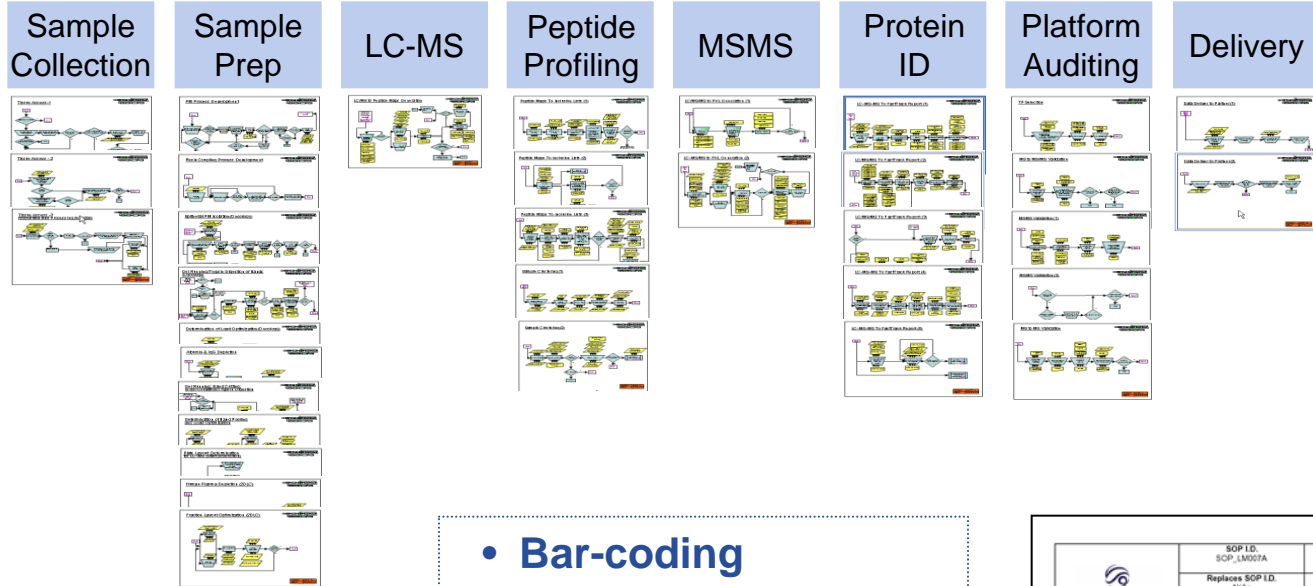
Targeted Sequencing

# QA ENVIRONMENT AND PROCESS ARE ESSENTIAL TO REDUCING VARIABILITY



CAPRION  
PROTEOMICS

## Gated Processes



Process Control

- Bar-coding
- Electronic Data Management (LIMS)
- Standard Operating Procedures
- Locked freezers
- Locked raw data
- Automatic backup

Chain of Custody

	SOP I.D. SCP_LM007A	CONFIDENTIAL
	Replaces SOP I.D. N/A	Author/Reviewer (Initials/Date)
<b>On-Line ZDLC-MS and ZDLC-MS/MS Sample Analysis by Micromass Ultima Q-ToF System</b>		
<b>1. Objective</b>		
This procedure describes the analysis of samples requiring on-line ZDLC-MS or ZDLC-MS/MS systems.		
<b>2. Purpose</b>		
The purpose of this procedure is to describe the methodology to be used for on-line ZDLC-MS and on-line ZDLC-MS/MS analysis of a sample using a Micromass Ultima Q-ToF LC/MS system.		
<b>3. Scope</b>		
This procedure applies to all samples requiring on-line analysis ZDLC-MS or ZDLC-MS/MS using Micromass Ultima Q-ToF LC/MS systems.		
<b>4. Responsibilities</b>		
4.1 LC-MS and LC-MS/MS Process Leader(s) are responsible for ensuring that all steps described within this procedure are followed and documented.		
4.2 All Users assigned to the sample analysis are responsible for following the steps described within this procedure.		
4.3 The Program Leader is responsible for determining the concentration of samples, volume to inject and the mode of injection.		
<b>5. Definitions and Acronyms</b>		
<b>5.1 Definitions</b>		
N/A		
Approved:	Dept:	Date:
Approved:	Dept:	Date:
Approved:	Dept: Quality Assurance	Date:
Effective Date: YYYYmmdd QA Final Approval		
<b>IT IS STRICTLY FORBIDDEN TO DUPLICATE THIS DOCUMENT</b>		
SOP N° 0106 2004.11.10 <span style="float: right;">Page 1 of 11</span>		

SOPs

## PROBLEM:

### No clear guidelines for the collection, processing, storage and analysis of plasma/serum for proteomics analysis

- Serum or plasma
- EDTA or heparin (for plasma)
- Added protease inhibitors (commercial tubes or DIY)
- Acceptable processing times and temperatures (before and after spin)
- Freezer temperature and storage time
- Multiple freeze-thaw cycles
- High abundance protein depletion

# SAMPLE COLLECTION AND STORAGE VARIABLES: FULL STUDY

## 1. Type of collection tube

- BD serum SST tube with gel and clot activator (red/grey top)
- BD heparin tube (green top)
- BD K<sub>2</sub>-EDTA tube (lavender top)
- Becton Dickinson (BD) P100
- BD K<sub>2</sub>-EDTA tube (lavender top) with protease inhibitor cocktail added at the time of pipetting separated plasma

## 2. Variation in the 2 key bench times

- Before centrifugation
- After centrifugation but before pipetting and freezing
- Up to 4 days
- 20°C or 37°C

## 3. Number of freeze-thaw cycles

## 4. Length of time in -20°C or -80°C storage

## 5. Cancer patients and age and gender-matched controls

- Prostate and breast cancer

# DESIGNING A WELL CONTROLLED STUDY

- Well defined and documented study design
- Protocols and SOPs for each step
- Bar coding of all tubes
- Chain of custody
- LIMS for information storage and sample tracking
- Well maintained and documented instrumentation
- Freezers on alarm with remote email alert
- Tool for managing blood collection, processing and storage
  - Interactive prompter and timer
  - Allows analysis of time taken for each step and variability
  - Allows documentation and comparison of process at multiple centers in clinical trial



# ORACLE-BASED LIMS (NAUTILUS)

Nautilus - limspord.labs.ppidi.local - [Exploring - P3874PAM001\_02PL]

File Edit View Window Help

P3874PAM001\_02PL

- + P0084
- + P0085
- + P0087
- + P0090
- + P1016
- + P1019
- + P1022
- + P3874
  - P3874PAM001
    - P3874PAM001\_01PL
    - P3874PAM001\_02PL
      - PLasmaTest
      - + P3874PAM001\_02PL01
      - + P3874PAM001\_02PL02
      - + P3874PAM001\_02PL03
      - + P3874PAM001\_02PL04
      - + P3874PAM001\_02PL05
      - + P3874PAM001\_02PL06
      - + P3874PAM001\_02PL07
      - + P3874PAM001\_02PL08
      - + P3874PAM001\_02PL09
    - P3874PAM001\_03PL
    - P3874PAM001\_04PL
    - P3874PAM001\_05PL
  - + P3874PAM002
  - + P3874PAM003
  - + P3874PAM004
  - + P3874PAM005

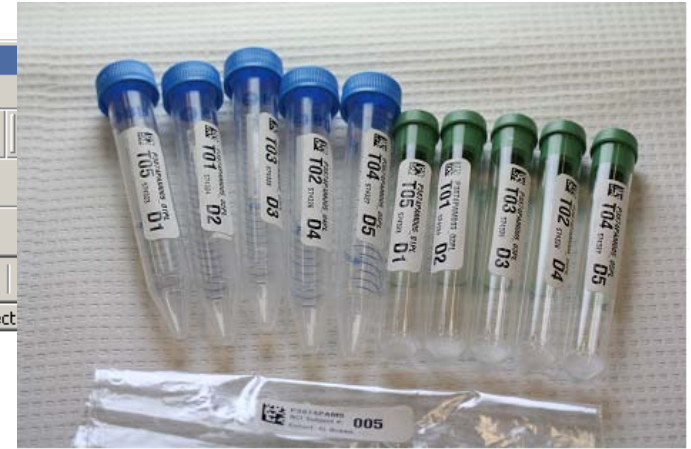
Name	Project
PLasmaTest	
P3874PAM001_02PL01	
P3874PAM001_02PL02	
P3874PAM001_02PL03	
P3874PAM001_02PL04	
P3874PAM001_02PL05	
P3874PAM001_02PL06	
P3874PAM001_02PL07	
P3874PAM001_02PL08	
P3874PAM001_02PL09	

Study

Subject

Draw/time-point

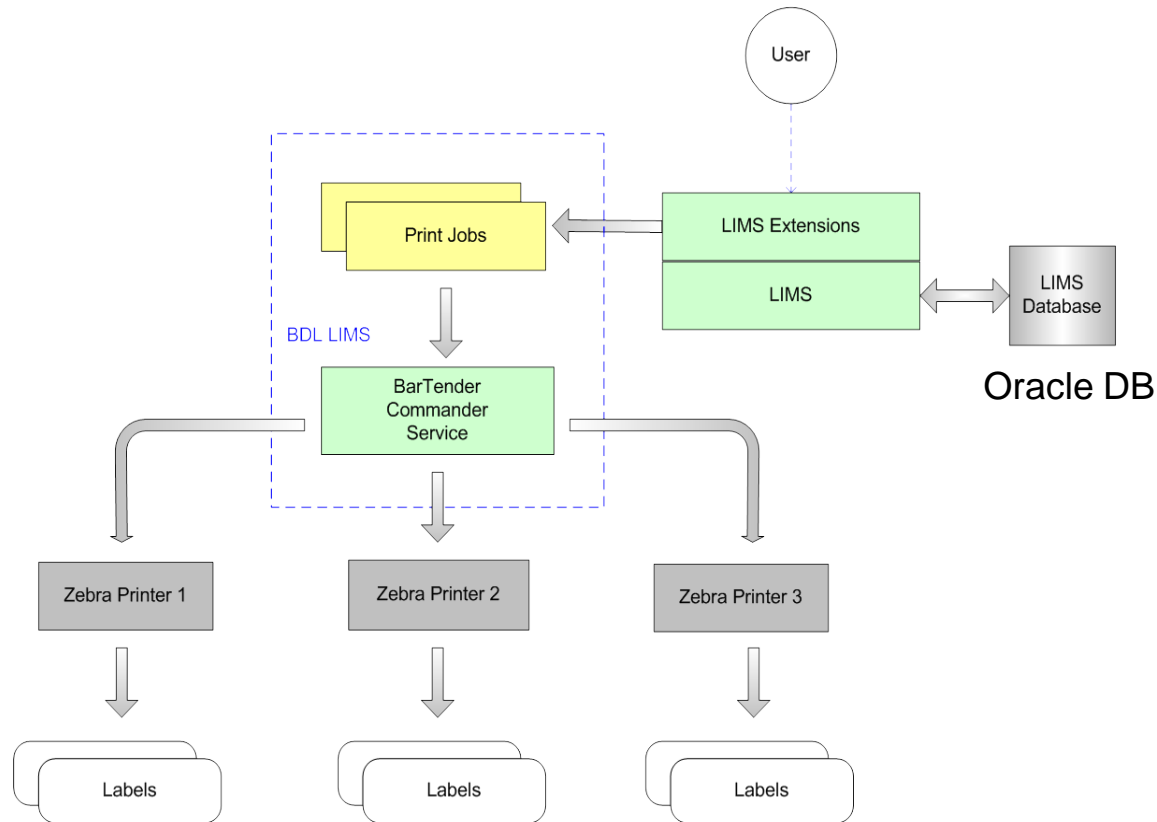
Frozen aliquot



574140

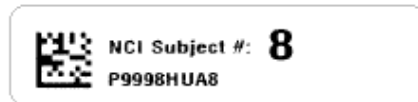


# LIMS Barcode Label System v2.0

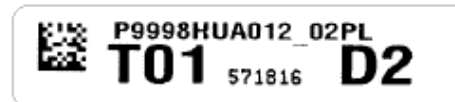


## Two-dimensional barcodes

Subject Label



Draw Label  
(5 per subject)

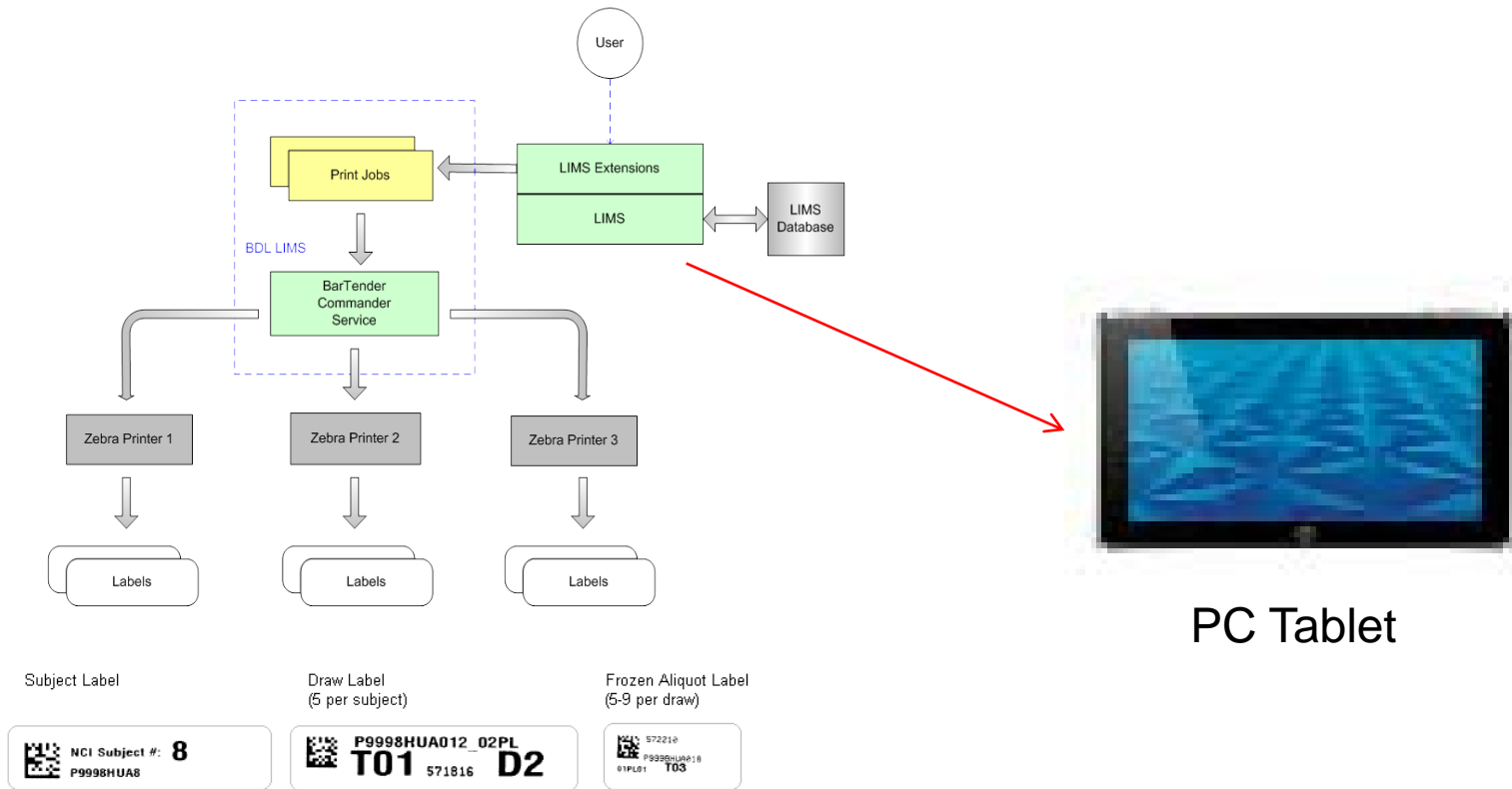


Frozen Aliquot Label  
(5-9 per draw)



# PC TABLET FOR PROCESS CONTROL

## LIMS Barcode Label System v2.0



# PC TABLET PROCESS CONTROL SCREEN



NCI Biospecimen Integrity Project - Sample Collector

Subject  
Sample  
Frozen Aliquot  
Notes

**Sample Tracking**    Tb Study 1 - Sample Collected - Patient Has Departed Current Time: **2:55:40 PM**

**Step 1: Arrange 5 draw tubes following this map, and scan draw barcodes on each tube:**

Pre-Label:	3	5	2	4	1
Tb #:	Tb 1	Tb 2	Tb 3	Tb 4	Tb 5
Draw Barcode:	<input type="text" value="571525"/>	<input type="text" value="571526"/>	<input type="text" value="571527"/>	<input type="text" value="571528"/>	<input type="text" value="571529"/>
	P9998HUA009_01P	P9998HUA009_02P	P9998HUA009_03P	P9998HUA009_04P	P9998HUA009_05P

**Step 2: Enter minutes since sample was drawn:**   Tb started at: **2:25:35 PM**

**Step 3: Enter Blood Level (cm):**

<input type="text" value="2.5"/>	<input type="text" value="3"/>	<input type="text" value="2.5"/>	<input type="text" value="2.5"/>	<input type="text" value="3"/>
Tb 1	Tb 2	Tb 3	Tb 4	Tb 5

**Tb Timing - Blood Draw to Centrifugation**

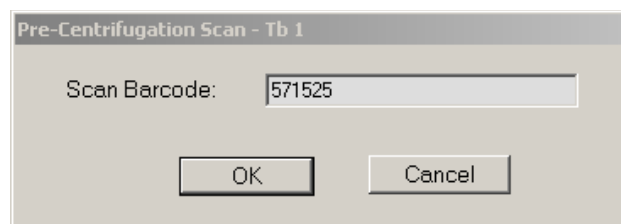
	Tb 1	Tb 2	Tb 3	Tb 4	Tb 5
Required Blood Time:	<input type="text" value="30 Minutes"/>	<input type="text" value="1 Hour"/>	<input type="text" value="2 Hours"/>	<input type="text" value="4 Hours"/>	<input type="text" value="6 Hours"/>
Count Down:	<b>00:04 OVER DUE</b>	<b>29:55</b>	<b>1:29:55</b>	<b>3:29:55</b>	<b>5:29:55</b>
When a timer is due (turns yellow), click button to proceed to centrifugation:	<input type="button" value="Start Centrif"/>	<input type="button" value="Start Centrif"/>	<input type="button" value="Start Centrif"/>	<input type="button" value="Start Centrif"/>	<input type="button" value="Start Centrif"/>

**Tp Timing Plasma Centrifugation to Pipetting**

	Tp 1	Tp 2	Tp 3	Tp 4	Tp 5
Required Plasma Time:	<input type="text" value="30 Minutes"/>	<input type="text" value="30 Minutes"/>	<input type="text" value="30 Minutes"/>	<input type="text" value="30 Minutes"/>	<input type="text" value="30 Minutes"/>
Count Down:	[Red]	[Red]	[Red]	[Red]	[Red]
When a timer is due (turns red), click button to proceed to Pipetting:	<input type="button" value="Start Pipetting"/>	<input type="button" value="Start Pipetting"/>	<input type="button" value="Start Pipetting"/>	<input type="button" value="Start Pipetting"/>	<input type="button" value="Start Pipetting"/>
	<input type="checkbox"/> Pipetting Done	<input type="checkbox"/> Pipetting Done	<input type="checkbox"/> Pipetting Done	<input type="checkbox"/> Pipetting Done	<input type="checkbox"/> Pipetting Done
	<input type="checkbox"/> Sample in Freezer	<input type="checkbox"/> Sample in Freezer	<input type="checkbox"/> Sample in Freezer	<input type="checkbox"/> Sample in Freezer	<input type="checkbox"/> Sample in Freezer

CS

2



**Click “Start Centrif” button,  
“Pre-Centrifugation Scan” pops up,  
requests to scan barcode on Vacutainer tube**

# Sample Page – Tp 1 time due, timer and “Start Pipetting” button turned red, flashes and emits repeating (distinct) warning tone



NCI Biospecimen Integrity Project - Sample Collector

**Sample Tracking**    Tb Study 1 - Sample Collected - Patient Has Departed
Current Time: **3:26:06 PM**

**Step 1: Arrange 5 draw tubes following this map, and scan draw barcodes on each tube:**

Pre-Label:	3	5	2	4	1
Tb #:	Tb 1	Tb 2	Tb 3	Tb 4	Tb 5
Draw Barcode:	571525	571526	571527	571528	571529
	P9998HUA009_01P	P9998HUA009_02P	P9998HUA009_03P	P9998HUA009_04P	P9998HUA009_05P

**Step 2: Enter minutes since sample was drawn:**  **And click:**       Tb started at: **2:25:35 PM**

**Step 3: Enter Blood Level (cm):**

	<input style="width: 60px;" type="text" value="2.5"/>	<input style="width: 60px;" type="text" value="3"/>	<input style="width: 60px;" type="text" value="2.5"/>	<input style="width: 60px;" type="text" value="2.5"/>	<input style="width: 60px;" type="text" value="3"/>
	Tb 1	Tb 2	Tb 3	Tb 4	Tb 5

**Tb Timing - Blood Draw to Centrifugation**

	Tb 1	Tb 2	Tb 3	Tb 4	Tb 5
Required Blood Time:	30 Minutes	1 Hour	2 Hours	4 Hours	6 Hours
Count Down:	<b>00:17 OVER DUE</b>	<b>00:07 OVER DUE</b>	<b>59:29</b>	<b>2:59:29</b>	<b>4:59:29</b>
When a timer is due (turns yellow), click button to proceed to centrifugation:	<input checked="" type="checkbox"/> Centrif Begun	<input checked="" type="checkbox"/> Centrif Begun	<input type="button" value="Start Centrif"/>	<input type="button" value="Start Centrif"/>	<input type="button" value="Start Centrif"/>

**Tp Timing Plasma Centrifugation to Pipetting**

	Tp 1	Tp 2	Tp 3	Tp 4	Tp 5
Required Plasma Time:	30 Minutes	30 Minutes	30 Minutes	30 Minutes	30 Minutes
Count Down:	<b>00:07 OVER DUE</b>				
When a timer is due (turns red), click button to proceed to Pipetting:	<input style="background-color: red; color: white;" type="button" value="Start Pipetting"/>	<input type="button" value="Start Pipetting"/>	<input type="button" value="Start Pipetting"/>	<input type="button" value="Start Pipetting"/>	<input type="button" value="Start Pipetting"/>
	<input type="checkbox"/> Pipetting Done	<input type="checkbox"/> Pipetting Done	<input type="checkbox"/> Pipetting Done	<input type="checkbox"/> Pipetting Done	<input type="checkbox"/> Pipetting Done
	<input type="checkbox"/> Sample in Freezer	<input type="checkbox"/> Sample in Freezer	<input type="checkbox"/> Sample in Freezer	<input type="checkbox"/> Sample in Freezer	<input type="checkbox"/> Sample in Freezer

CS

# “Deviation Report” Page



NCI Biospecimen Integrity Project - Sample Collector

Subject  
Sample  
Frozen Aliquot  
Notes

Any Deviations from the SOP?

Any Unusual Observations about the Subject?

Any Unusual Observations about the venipuncture collection?

Any Unusual Appearance or Behavior of the Blood or Plasma/Serum?

Complete

102

# LIMS METADATA STORAGE (ELECTRONIC MEDICAL RECORD)



**Medical Record**

Enter subject #:  Subject ID:

Signed Informed Consent?  Yes  No Year of Birth:  Gender:  Male  Female Approx Date of Initial Cancer Diagnosis:

Current Cancer Treatments (last 3 months):  
 Chemotherapy, Oral  
 Chemotherapy, IV or IM (by needle)  
 Radiation  
 None

Most recent one or few blood tests, especially capture data on cancel markers if recent (last 6 months)

Recent Medications:

Other Active Health Problems:

Breast Cancer Info:  
 Surgery, Lumpectomy Date:   
 Surgery, Mastectomy Date:   
Menopause Status:   
Estrogen Receptor (ER) Status:   
Progesterone Receptor (PR) Status:   
HER2/ERBB2 IHC Status:   
Histological Type:

Prostate Cancer Info:  
 Surgery, removal of prostate Date:   
TNM Staging System Numbers: T=  N=  M=   
Gleason grade:   
Gleason score:   
Other notable comments:

Tumor Stage:  Coordinator initials:  Date:



# Preliminary Biospecimen Analysis

# PRELIMINARY BIOSPECIMEN ANALYSIS

## Scope of preliminary study:

- Proteomic analysis on plasma from cancer patients
  - 3 tube types:
    - Heparin, EDTA, EDTA/PI
  - 3 incubation times at RT prior to centrifugation
    - 0.5, 4 and 24 h
  - n = 10

## Analysis performed:

- Samples depleted of high abundance plasma proteins with Agilent MARS-14 column
- Protein digested with trypsin and analyzed by LC-MS
- Peak alignment and matching performed with Rosetta's Elucidator software
- Differential expression analysis
- Peptides sequenced and clustered

# PEPTIDE POPULATION DIFFERS BETWEEN HEPARIN AND EDTA PLASMA

## Heparin vs. EDTA

- Total components: 6,600
- Heparin-specific components: 5,000
- EDTA+/-PI-specific components: 5,700
- Only approx. 4,000 (61%) components shared between heparin and EDTA
- 83% proteins are shared between EDTA and heparin
- Total number of proteins detected is almost the same

## EDTA +/- Protease Inhibitors

- 140/5700 (2.5%) components differ > 2 fold between EDTA +/- PI
- Adding protease inhibitors increases number of proteins detectable from 135 to 137

# FEW CHANGES SEEN OVER TIME

Comparison ID	Description	Differentially expressed peptides	Differentially expressed proteins
1	[Heparin] 0.5 hr vs 4 hr	0	0
2	[Heparin] 0.5 hr vs 24 hr	2	1
4	[EDTA] 0.5 hr vs 4 hr	3	3
5	[EDTA] 0.5 hr vs 24 hr	11	4
7	[EDTA+PI] 0.5 hr vs 4 hr	4	4
8	[EDTA+PI] 0.5 hr vs 24 hr	12	5

## Selection criteria:

- Fold-change  $\geq 2$ ;
- p-value  $\leq 0.05$ ;
- q-value  $\leq 0.05$

## Results:

- Small effects of incubation time
- Only slightly more changes with EDTA than heparin
- Small effect of adding protease inhibitors

# IMPACT OF SMALLER CHANGES ON CALCULATING DIFFERENTIALLY EXPRESSED PEPTIDES



Comparison_ID	Description	DI > 2 pvalue <= 0.05 qvalue <= 0.05	DI > 1.5 pvalue <= 0.05 qvalue <= 0.05
1	[Heparin] 0.5 hr vs 4 hr	0	1
2	[Heparin] 0.5 hr vs 24 hr	3	3
3	[Heparin] 4 hr vs 24 hr	2	3
4	[EDTA] 0.5 hr vs 4 hr	3	11
5	[EDTA] 0.5 hr vs 24 hr	21	27
6	[EDTA] 4 hr vs 24 hr	20	29
7	[EDTA+PI] 0.5 hr vs 4 hr	7	12
8	[EDTA+PI] 0.5 hr vs 24 hr	26	33
9	[EDTA+PI] 4 hr vs 24 hr	21	30
10	[0.5 hr] Heparin vs EDTA	2,606	3,569
11	[0.5 hr] Heparin vs EDTA+PI	2,897	3,804
12	[0.5 hr] EDTA vs EDTA+PI	141	346
13	[4 hr] Heparin vs EDTA	2,472	3,408
14	[4 hr] Heparin vs EDTA+PI	2,785	3,656
15	[4 hr] EDTA vs EDTA+PI	77	178
16	[24 hr] Heparin vs EDTA	2,729	3,642
17	[24 hr] Heparin vs EDTA+PI	2,846	3,765
18	[24 hr] EDTA vs EDTA+PI	40	76
19	0.5 hr vs 4 hr	2	7
20	0.5 hr vs 24 hr	19	21
21	4 hr vs 24 hr	20	24
22	Heparin vs EDTA	2,732	3,778
23	Heparin vs EDTA+PI	2,967	3,971
24	EDTA vs EDTA+PI	140	368
		3,386	4,498

# OXIDIZED PEPTIDES ARE NOT SELECTIVELY DISTRIBUTED

Differentially expressed components/peptides/proteins

(DI > 2 | pvalue <= 0.05 | qvalue <= 0.05)

Comparison_ID	Description	#Components	#Peptides with oxidation PTM		#Proteins with oxidation PTM	
			Upregulated	Downregulated	Upregulated	Downregulated
19	0.5 hr vs 4 hr	2	0	0	0	0
20	0.5 hr vs 24 hr	19	0	0	0	0
21	4 hr vs 24 hr	20	0	0	0	0
22	Heparin vs EDTA	2,732	13	34	10	6
23	Heparin vs EDTA+PI	2,967	11	35	8	7
24	EDTA vs EDTA+PI	140	0	0	0	0
		3,386	17	36	13	8

# SEMI-TRYPTIC AND NON-TRYPTIC PEPTIDES ACROSS THREE STUDIES

	NCI	NIAID site 1	NIAID site 2
# Sequenced Peptides	3,158	1,527	2,593
# (Fully) Non-tryptic	11	17	1
# Semi-tryptic	506	521	377
# Tryptic	2,641	989	2,215
% semi or non-tryptic peptides	16%	35%	15%

## CONCLUSIONS OF PRELIMINARY STUDY

- Large number of components detectable in only heparin or EDTA tubes
- 14% more components detectable in EDTA tubes
- Very few components changed in concentration in any tube type after 24h at RT
- Little effect of protease inhibitor cocktail in EDTA tubes after 24h at RT
- The median CV for normalized peptide intensities within each group is very low at 5-7% (mainly processing related)
- Non-normalized raw intensity median CVs ~ 32-40% (processing and biological variability)
- 16% of sequenced peptides were cleaved at a site other than trypsin, suggesting some degradation
  - Similar to one study (well managed samples)
  - Better than a second study (older sample set, multiple freeze-thaws)



# QUALITY ASSESSMENT ASSAYS

- Multiplexed ELISA assays: Luminex
- Multiplexed MRM mass spec assays

# INDEPENDENT ASSESSMENT OF PROTEIN CONCENTRATION AND VARIATION

Lower abundance protein panel:

IL6  
CA125  
CA19-9  
MUC1  
PSA  
PRL  
LEP  
OPN  
MIF  
AFP  
CEA

Higher abundance protein panel:

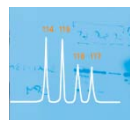
VTN  
ECM1  
F13A  
VDP  
AT3  
CFH  
FCN3  
LUM

\* This group will also be measured by LC-MS

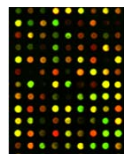
# MULTIPLEXED MRM ASSAY DEVELOPMENT STRATEGY



1. Literature



2. Proteomics

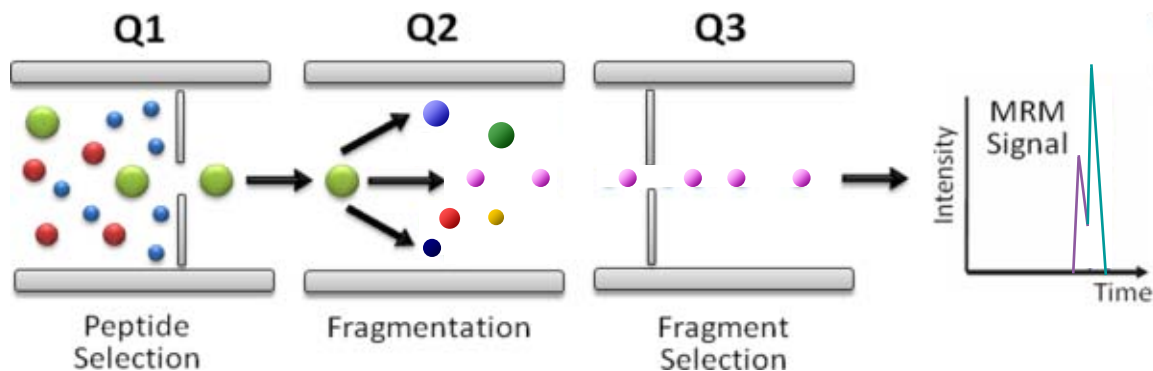


3. Transcript Profiling

## Protein Sequence

MSAIQAAWPSGTECIAKYNFHGTAEQD  
LPFCK**GDVLTIVAVTK**DPNWKAKNKV  
GREGIIPANYVQKREGVKAGTKLSLMP  
WFHGKITREQAER**LLYPPETGLFLVRE**  
STNYPGDYTLVSCDVGKVEHYRIMYHA  
SKL**SIDEEVYFENL**KMQLVEHYTSDAD  
GLCTRLIKPKVMEGTVAQAQDEFYRSGW  
ALNMKELKLLQTIGK**GFEGDVMLGDYR**  
GNKVAVKCIKNDATA...

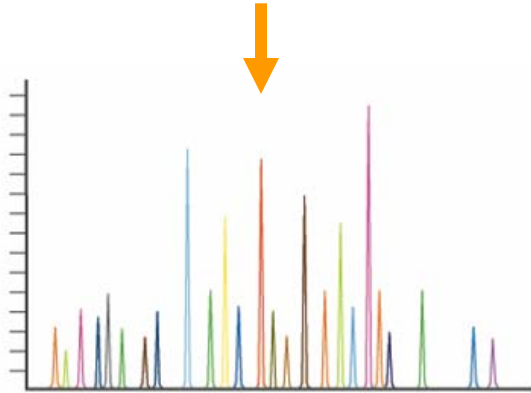
Candidate markers for MRM assay development can come from proteomics or other sources, including the literature



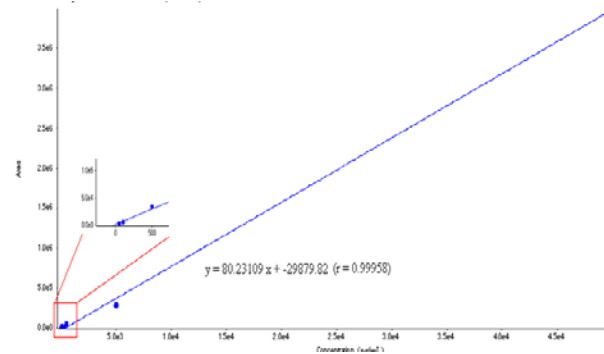
Double selection improves signal/noise and reduces interference

# MRM-MS ASSAYS: VERIFY AND VALIDATE CANDIDATE BIOMARKERS

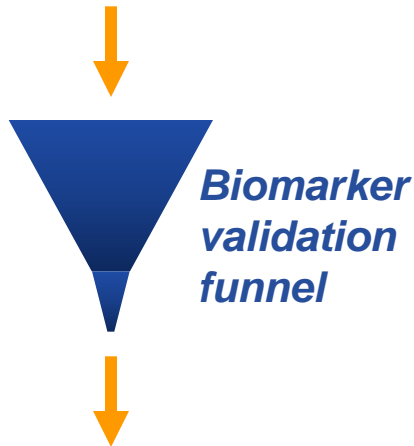
Candidate Biomarkers



Multiplexed MRM assay  
1-700 candidate biomarkers



Linear and quantitative

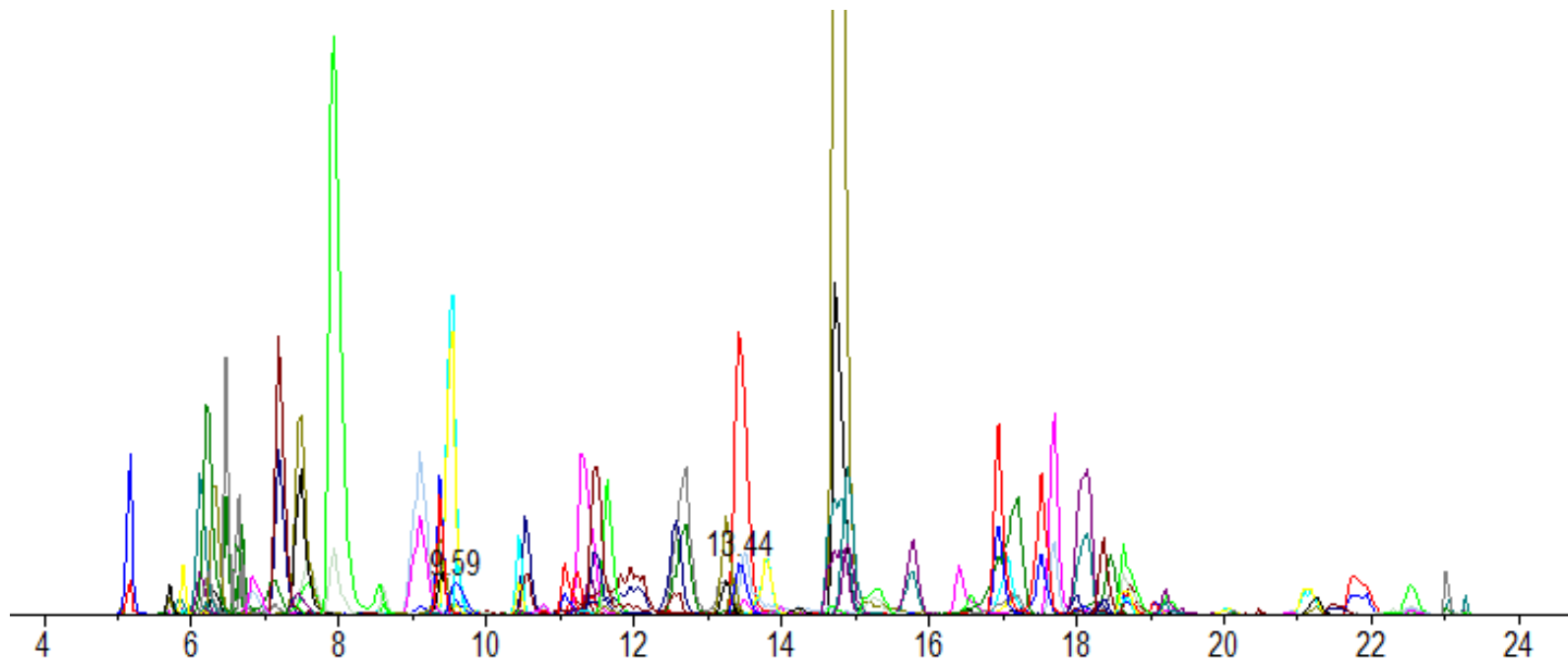


Validated biomarkers for  
MRM or ELISA assay

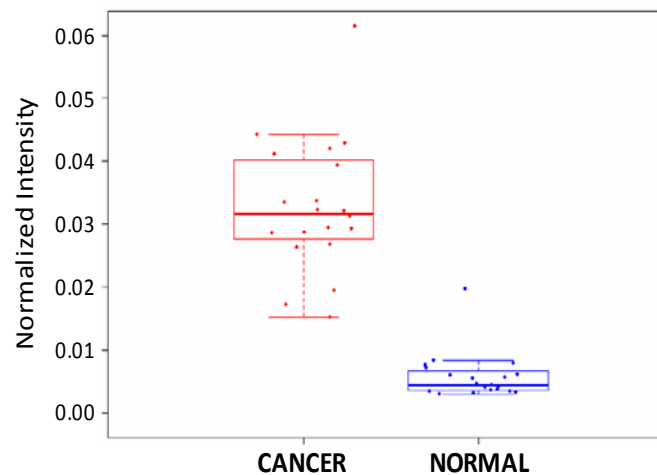
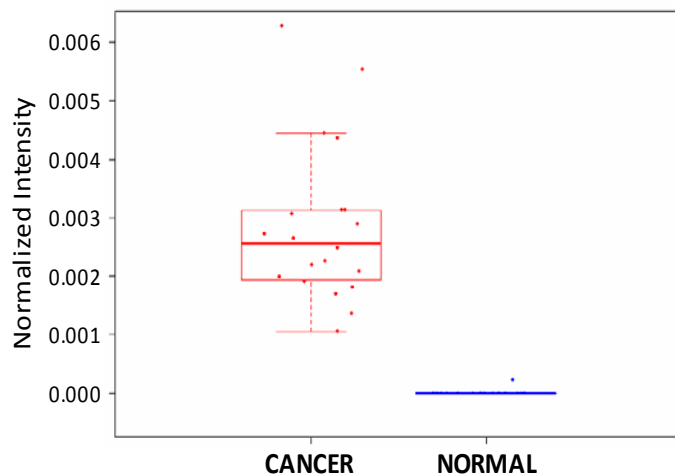
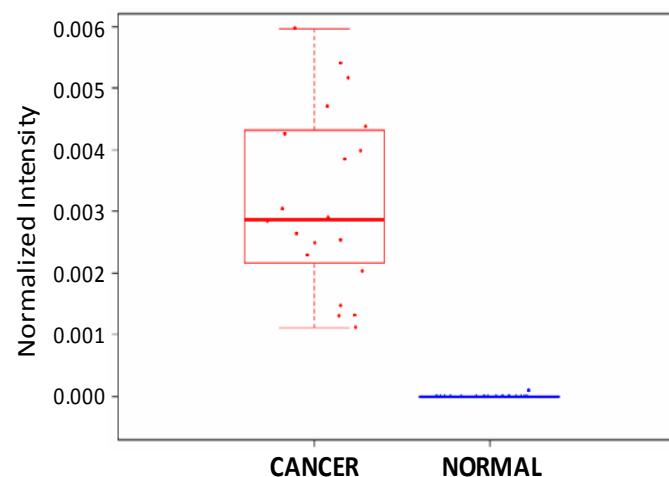
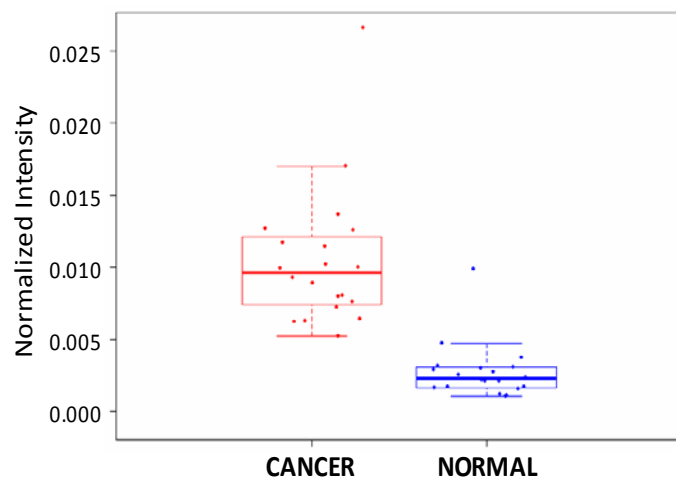
- Rapid assay development
- Multiplexed (up to 700 biomarkers)
- Confirms biomarker ID
- Confirms differential expression
- Determines absolute abundance
- Can be validated for regulatory compliance

# EXAMPLE OF MRM ASSAY DEVELOPED FROM LIST OF 90 LUNG CANCER Dx CANDIDATES

- Predict best 5 peptides/protein, synthesize
- Determine empirically the best 2, monitor 2 transitions each
- Pre-verification study with plasma from 20 cancer and 20 control subjects
- 64 of the targeted proteins (71%) successfully detected in un-spiked plasma samples (cancer and controls)



# CANDIDATE CLASSIFIERS IDENTIFIED



Current study: Larger set of proteins (700 candidate markers) and more samples (~400) from multiple sources

